



ARTICLE

Enriched Artemia Nauplii with Commercial Probiotic in the Larviculture of Angelfish *Pterophyllum scalare* Lichtenstein (1823)

Natalino da Costa Sousa^{1*} José Araújo da Silva¹ Emily Monteiro Lopes¹ Arthur Felipe Lima dos Santos¹ Francisco Alex Lima Barros¹ Carlos Alberto Martins Cordeiro¹ Peterson Emmanuel Guimarães Paixão² Estela dos Santos Medeiros³ João Carlos Nunes de Souza³ Márcia Valéria Silva do Couto²

1. Federal University of Pará (UFPA), Campus Bragança, Alameda Leandro Ribeiro, Bragança, 68600-000, PA, Brazil
2. Tiradentes University (UNIT) - Av. Murilo Dantas, 300 - Farolândia, CEP 49032-490, Aracaju, Sergipe, Brazil
3. Faculdade Pio Décimo Av. Pres. Tancredo Neves, 5655 - Jabotiana, 49095-000, Aracaju – SE. Brazil

ARTICLE INFO

Article history

Received: 17 December 2019

Accepted: 31 December 2019

Published Online: 28 February 2020

Keywords:

Ornamental fish

Probiotic

Larviculture

ABSTRACT

This study evaluated the effect of enriched artemia nauplii with commercial probiotic for angelfish larvae determining growth performance, survival and modulation of the intestinal microbiota. Therefore, it experiment occurred in completely randomized design with five treatments (T1- 0.0, T2- 1.5, T3- 3.0, T4- 4.5 and T5- 6.0g of commercial probiotic) and four replaces during 20 days. After larviculture, post larvae passed by biometric procedures to determine productive performance and then microbiological analysis. Occurred reduction of total heterotrophic bacteria while increased lactic acid bacteria in the intestinal tract from the post larvae for treatments T3, T4 and T5. The commercial probiotic also increased the survival and performance as final weight, weight gain and specific growth rate. For these reasons, the use of 3g of commercial probiotic promotes greater performance and intestinal modulation for angelfish larvae.

1. Introduction

Ornamental fish trade around the world has become a profitable activity moving approximately US 15 million ^[1,2]. Currently, it market look for several fish species with highest quality, different shape and colors ^[3,4]. Among the native fish species from Amazon, the angelfish *Pterophyllum scalare* has economic potential into national and international market due to this varied patterns and colors required by the ornamental market ^[4,5,6].

In freshwater ornamental fish, the larviculture remains as the most problematic phase of production with highest mortality rate caused by management, inadequate nutrition and stocking density ^[4,7,8]. These factors can reduces its productive performance and health of the larvae ^[4,5,9].

The larval phase, live feed have an important paper to supply the nutritional requirement and its development ^[8]. Currently, the ornamental fish sector use widely artemia nauplii due to the size, protein and lipid profile as well as enzymes that aid its digestion process ^[10,11]. In the last de-

*Corresponding Author:

Natalino da Costa Sousa,

Federal University of Pará (UFPA), Campus Bragança, Alameda Leandro Ribeiro, Bragança, 68600-000, PA, Brazil;

Email: natal.engpesca@gmail.com

cade, fish farmers has applied different strategies for larvaculture to improve the performance and health [8,12,13,14]. Among the new strategies, enriched artemia nauplii with probiotic showed nice results about the productive performance, survival, improvement of immunological system and intestinal modulation [9,15,16,17,18,19].

In front this, enriched diets for ornamental fish larvae becomes an efficient strategy in captivity rearing. Nonetheless, still missing scientific data about the use of probiotic for larval diet. Thus, this study evaluated the effect of enriched artemia nauplii with commercial probiotic for angelfish larvae *Pterophyllum scalare* determining performance, survival and intestinal modulation.

2. Material and Method

2.1 Experimental Design

This study used larvae of angelfish *Pterophyllum scalare* (1.10±0.01 mg and 4.70±0.25 mm) from natural reproduction, placed 10 fish per polyethylene tanks (total capacity for one liter). Therefore, it experiment occurred in completely randomized design with five treatments (C- 0.0, T1- 1.5, T2- 3.0, T3- 4.5 and T4- 6.0 g of commercial probiotic) and four replaces during 20 days.

The feeding management have four daily frequencies (08, 11, 14 and 17 hours) providing one hundred artemia nauplii per larvae [5]. After the last daily feeding, it was carried out water exchange (30%). The water quality parameters were: temperature (28.6±0.35 °C), dissolved oxygen (5.89±0.11 mg.L⁻¹) pH (6.56±0.42) total ammonia (0.18±0.04 mg.L⁻¹) and electric conductivity (168±28.32 μS.cm⁻¹).

2.2 Biological Culture and Enrichment Process

To obtain the artemia nauplii, it used cyst 1g into the tank (1 liter) with constant aeration during 24 hours [9]. After hatching time, artemia nauplii placed in becker (50mL) received commercial probiotic *Lactobacillus acidophilus* (1.1 x 10⁸ CFU.g⁻¹) and *Enterococcus faecium* 7.7x 10⁷ CFU.g⁻¹ during 40 minutes before feeding management [20].

2.3 Intestinal Microbiota

For determine bacterial amount in the intestine from post larvae, its intestinal tract (pools of five larvae) macerated with sterile saline solution 0.65% passed by serial dilution (1:10 factor). An aliquot (100 μL) from each of three dilutions (10⁻¹, 10⁻³ and 10⁻⁵) was used to inoculate petri dishes containing either Triptone Soy Agar (TSA - incubated at 30 °C for 24 hours) to obtain counts of total heterotrophic bacteria, or MRS Agar (incubated at 30 °C for

48 hours) to obtain counts of lactic acid bacteria [19].

2.4 Growth Performance

At the end of experiment, all post larvae passed per biometric procedure to determine weight and length evaluating: Total length (TL), final weight (FW), weight gain (WG), specific growth rate (SGR), survival (S), uniformity (U) [21] and relative condition factor (Kr) [22].

2.5 Statistical Analysis

Data was tested for normality (Shapiro-Wilk) and homoscedasticity (Levene's). Subsequent analysis of variance tests were performed applying post hoc Tukey tests for separation of means (P<0.05) [23]. Microbiological counts showed non-parametric characteristics and were therefore log transformed [$\log_{10}(x+1)$] and arc sen square root (x).

3. Result

At the end of experiment, occurred reduction (p<0,05) of total heterotrophic bacteria (T2= 3.41±0.061; T3= 3.43±0.078 and T4= 3.36±0.04 Log CFU.g⁻¹), while increased (p<0,05) lactic acid bacteria in the intestinal tract from the post larvae for treatments T2 (3.81±0.06 Log CFU.g⁻¹), T3 (3.95±0.08 Log CFU.g⁻¹) and T4 (3.95±0.12 Log CFU.g⁻¹), compared to control treatments (Figure 1).

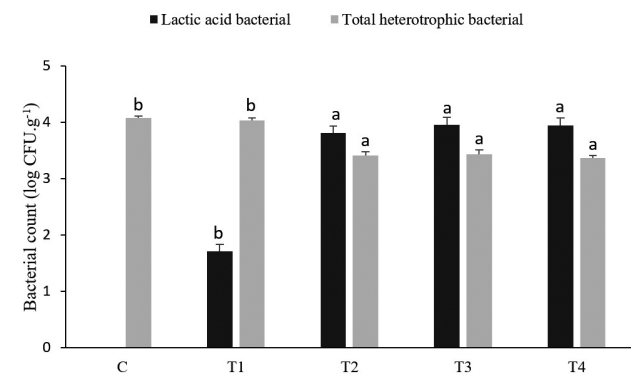


Figure 1. Heterotrophic and lactic acid bacterial counting from intestinal tract of angelfish larvae *Pterophyllum scalare*

Note: Different letters mean statistical difference (p<0.05) among the treatments.

After 20 days, With regard to the productive performance, larvae fed with the enriched artemia nauplii with T2, T3 and T4 treatment, increased (p<0,05) final weight (FW), weight gain (WG) and specific growth rate (SGRw) for when compared to the control (table 1). Furthermore, larvae mortality significantly increased (p<0,05) in the control (72,0±8,5) (table 1).

Table 1. Productive performance and survival from angelfish larvae *Pterophyllum scalare* fed with *Artemia* sp. without enrichment (C) and enriched with commercial probiotic (T1: 1.5 g; T2: 3.0g; T3: 4.5g and T4: 6.0g) during larviculture

Treatments	TL (mm)	FW (mg)	WG (mg)	SGR _w (%)	SGR _L (%)	UNI (%)	Kr	S (%)
C	12,48±0,35 a	38±1,1 b	35±1,1 b	23,2±0,17 b	5,74±0,03 a	66,0±16,2 a	0,99±0,02 a	72,0±8,5 b
T1	12,47±0,32 a	39±1,2 b	35±1,2 b	23,4±0,18 b	5,71±0,16 a	75,8±12,4 a	0,99±0,02 a	84,5±12,5 a
T2	12,83±0,24 a	42±1,3 a	39±1,8 a	24,8±0,32 a	5,90±0,20 a	78,1±15,0 a	1,00±0,01 a	92,0±9,4 a
T3	12,86±0,48 a	41±1,7 a	40±2,1 a	25,0±0,47 a	5,77±0,23 a	76,2±14,8 a	0,99±0,01 a	95,5±8,8 a
T4	12,71±0,18 a	42±1,9 a	39±1,6 a	25,2±0,53 a	5,83±0,23 a	75,0±12,8 a	0,99±0,02 a	94,0±8,2 a

Note: Mean values ± standard deviation from productive performance, different letters in column mean statistical difference (p<0.05).

4. Discussion

The use of bacteria with probiotic potential has showed positive results to the ornamental aquaculture^[9,17,24,25]. Nonetheless, still missing reports about their use for angelfish larvae on productive performance, intestinal modulation and survival.

Colonize the intestinal tract stands out as the main aspect to determine its probiotic potential^[19,26]. For this study, the artemia nauplii works as transporter of probiotic bacterium to the host. Its intestinal tract reduced the heterotrophic bacteria amount with the increases of probiotic bacterium due to the competition for space, nutrients and releasing of bactericides^[15,19,27].

^[17] reported it modulator effect in the intestine for angelfish adult, as also to *Carassius auratus*^[24] and *Xiphophorus helleri*^[25]. In addition, probiotic act as immune stimulant forming barriers against pathogenic bacteria and increasing its defense cell numbers^[16,19,25,28]. Thus, it increases above 80% of survival could be related to the effects of probiotic bacterium included in the exogenous diet during larviculture.

The larval performance improved in this study with the enrichment of artemia nauplii promoting greater weight gain, specific growth rate and survival. It benefits would be related to the intestinal modulation that provides increased intestinal villi and better nutrient absorption^[9,19]. According to^[29], they observed similar result with increased performance for angelfish larvae *Pterophyllum scalare* using a different commercial probiotic.

For these reasons, enrichment of artemia nauplii becomes an efficient strategy to ornamental fish farming considering the probiotic amount to reach its benefits results. However, its microorganisms could influenced by environmental factors, age, development phase and physiological aspects^[19,30,31,32].

5. Conclusion

The enriched artemia nauplii with commercial probiotic (3g/L) can modulates the intestinal tract, increases the per-

formance and survival. Thus, it enriched diet can be used as a new strategy for larviculture of the angelfish larvae *Pterophyllum scalare*.

Conflicts of interest

The authors have no conflicts of interest to declare.

Reference

- [1] Zuanon, J.A.S.; Salario, A.L.; Furuya, W.M. Produção e nutrição de peixes ornamentais. Revista Brasileira de Zootecnia, 2011, 40: 165-174.
- [2] FAO – Food And Agriculture Organization of the United Nations. FAO Aquacultura Newsletter. Nº 56 (April), 2017: 64.
- [3] Abe, H.A., DIAS, J.A.R., Cordeiro, C.A.M., Ramos, F.M., Fujimoto, R.Y. *Pyrrhulina brevis* (steindachner, 1876) como uma nova opção para a piscicultura ornamental nacional: larvicultura. Boletim do Instituto de Pesca, 2015, 41: 113-122.
- [4] Abe HA, Dias J.A, Reis R.G., Sousa N.C., Ramos, F.M, Fujimoto, R.Y. Manejo alimentar e densidade de estocagem na larvicultura do peixe ornamental amazônico *Heros severus*. Boletim do Instituto de Pesca, 2016, 42: 514-22.
- [5] Pereira, S.L., Gonçalves-Júnior, L.P., Azevedo, R. Diferentes estratégias alimentares na larvicultura do acará-bandeira (*Pterophyllum scalare*, Cichlidae). Acta Amazonica, 2016, 46: 91-98. DOI: 10.1590/1809-4392201500472
- [6] Fujimoto, R.Y., Santos, R.F.B., Dias, H.M., Ramos, F.M., Silva, D.J.F., Honorato, C. A. Feeding frequency on the production viability of production and quantitative descriptors of parasitism in angelfish. Ciência Rural, 2016, 46: 304-309. DOI: 10.1590/0103-8478cr20141704
- [7] Gonçalves-Junior, L. P., Mendonça, P.P., Pereira, S.L., Matielo, M.D., AMORIM, I.R.S. Densidade de estocagem durante a larvicultura do kinguio. Boletim do Instituto de Pesca. 2016, 40: 597-604.
- [8] Couto, M.V.S.D., Sousa, N.D.C., Abe, H.A., Dias,

- J.A.R., Meneses, J.O., Paixão, P.E.G., Cunha, F.S., Ramos, F.M., Maria, A.N., Carneiro, P.C.F., Fujimoto, R. Y. Effects of live feed containing *Panagrellus redivivus* and water depth on growth of *Betta splendens* larvae. *Aquaculture research*, 2018, 49: 2671-2675.
DOI: 10.1111/are.13727
- [9] Azevedo, R.V., Fosse-Filho, J.C., Pereira, S.L., Andrade, D.R., Júnior, V.M. Prebiótico, probiótico e simbiótico para larvas de *Trichogaster leeri* (Bleeker, 1852, Perciformes, Osphronemidae). *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*, 2016, 68: 795-804.
DOI: 10.1590/1678-4162-8580
- [10] Malla, S., Banik, S. (2015). Production and application of live food organisms for freshwater ornamental fish Larviculture. *Advances in BioResearch*, 6: 159–167.
- [11] Loh, J.Y., Ting, A.S.Y. Effects of potential probiotic *Lactococcus lactis* subsp. *lactis* on digestive enzymatic activities of live feed *Artemia franciscana*. *Aquaculture international*, 2016, 24: 1341-1351.
DOI: 10.1007/s10499-016-9991-2
- [12] Abe, H.A., Dias, J.A.R., Sousa, N.D.C., Couto, M.V.S.D., Reis, R.G.A., Paixão, P.E.G., Fujimoto, R.Y. Growth of Amazon ornamental fish *Nannostomus beckfordi* larvae (Steindachner, 1876) submitted to different stocking densities and feeding management in captivity conditions. *Aquaculture Research*, 2019, 50: 2276-2280.
DOI: 10.1111/are.14108
- [13] Fabregat, T.E.H.P., Wosniak, B., Takata, R., Miranda Filho, K.C., Fernandes, J.B.K., Portella, M.C. Larviculture of siamese fighting fish *Betta splendens* in low-salinity water. *Boletim do Instituto de Pesca*, 2017, 43: 164-171.
DOI: 10.20950/1678-2305.2017v43n2p164
- [14] Ahmadifard, N., Aminloo, V. R., Tukmechi, A., Agh, N. Evaluation of the impacts of long-term enriched *Artemia* with *Bacillus subtilis* on growth performance, reproduction, intestinal microflora, and resistance to *Aeromonas hydrophila* of ornamental fish *Poecilia latipinna*. *Probiotics and antimicrobial Proteins*, 2018, 11: 957–965.
DOI: 10.1007/s12602-018-9453-4
- [15] Mouriño, J.L.P., Vieira, F.D., Jatobá, A.B., Silva, B.C., Jesus, G.F.A., Seiffert, W.Q., Martins, M.L. Effect of dietary supplementation of inulin and *W. cibaria* on haemato-immunological parameters of hybrid surubim (*Pseudoplatystoma* sp). *Aquaculture Nutrition*, 2012, 18: 73-80.
DOI:10.1111/j.1365-2095.2011.00879.x
- [16] Mouriño, J.L.P., Vieira, F.N., Jatobá, A., Silva, B.C., Pereira, G.V., Jesus, G.F.A., Martins, M.L. Symbiotic supplementation on the hemato-immunological parameters and survival of the hybrid surubim after challenge with *Aeromonas hydrophila*. *Aquaculture Nutrition*, 2015, 23: 276-284.
DOI: 10.1111/anu.12390
- [17] Azimirad, M., Meshkini, S., Ahmadifard, N., Hoseinifar, S. H. The effects of feeding with synbiotic (*Pediococcus acidilactici* and fructooligosaccharide) enriched adult *Artemia* on skin mucus immune responses, stress resistance, intestinal microbiota and performance of angelfish (*Pterophyllum scalare*). *Fish & shellfish immunology*, 2016, 54: 516-522.
DOI: 10.1016/j.fsi.2016.05.001
- [18] Vázquez-Silva, G., Ramírez-Saad, H.C., Aguirre-Garrido, J. F., Mayorga-Reyes, L., Azaola-Espinosa, A., Morales-Jiménez, J. Effect of bacterial probiotics bio-encapsulated into *Artemia franciscana* on weight and length of the shortfin silverside (*Chirostoma humboldtianum*), and PCR-DGGE characterization of its intestinal bacterial community. *Latin american journal of aquatic research*, 2017, 45: 1031-1043.
DOI: 10.3856/vol45-issues5-fulltext-18
- [19] Sousa, N. C, do Couto, M. V. S., Abe, H. A., Paixão, P. E. G., Cordeiro, C. A. M., Monteiro Lopes, E., Ready, J. S., Jesus, G. F. A., Martins, M. L., Mouriño, J. L. P., Carneiro, P. C. F., Maria, A. N., & Fujimoto, R. Y. (2019). Effects of an *Enterococcus faecium* based probiotic on growth performance and health of *Pirarucu*, *Arapaima gigas*. *Aquaculture Research*, 50, 3720–3728.
<https://doi.org/10.1111/are.14332>
- [20] Vázquez-Silva, G., Castro-Mejía, J.J., Sánchez de la Concha, B., González-Vázquez, R., Mayorga-Reyes, L., Azaola-Espinosa, A. Bioencapsulation of *Bifidobacterium animalis* and *Lactobacillus johnsonii* in *Artemia franciscana* as feed for charal (*Chirostoma jordani*) larvae. *Revista Mexicana de Ingeniería Química*, 2016, 15: 809-818.
- [21] Furuya, W.M., Souza, S.R.D., Furuya, V.R.B., Hayashi, C. Ribeiro, R.P. Pelletized and extruded diets for reversed Nile tilapia (*Oreochromis niloticus* L.) males, in finishing phase. *Ciência Rural*, 1998, 28, 483–487.
DOI:10.1590/S0103-84781998000300022
- [22] Le Cren, E. D. The length-weight relationship and seasonal cycle in gonad weight and condition in the perch (*Perca fluviatilis*). *Journal of Animal Ecology*, 1951, 20: 201- 219.
- [23] Zar, J. H. *Biostatistical Analysis*. 5th. Edition, Pren-

- tice-Hall, Inc., Upper Saddle River, New Jersey, USA. 2009: 576.
- [24] Mehdinejad, N., Imanpour, M.R., Jafari, V. Combined or Individual Effects of Dietary Probiotic *Pedococcus acidilactici* and Nucleotide on Growth Performance, Intestinal Microbiota, Hemato-biochemical Parameters, and Innate Immune Response in Goldfish (*Carassius auratus*). *Probiotics and antimicrobial proteins*, 2017, 10: 558–565.
DOI: 10.1007/s12602-017-9297-3
- [25] Hoseinifar, S.H., Roosta, Z., Hajimoradloo, A., Vakil, F. The effects of *Lactobacillus acidophilus* as feed supplement on skin mucosal immune parameters, intestinal microbiota, stress resistance and growth performance of black swordtail (*Xiphophorus helleri*). *Fish & shellfish immunology*, 2015, 42: 533-538.
DOI:10.1016/j.fsi.2014.12.003
- [26] Feng, J., Li, D., Liu, L., Tang, Y., Du, R. Characterization and comparison of the adherence and immune modulation of two gut *Lactobacillus* strains isolated from *Paralichthys olivaceus*. *Aquaculture*, 2019, 499: 381-388.
DOI: 10.1016/j.aquaculture.2018.08.026
- [27] Balcázar, J.L., Vendrell, D., Blas, I., Ruiz-Zarzuela, I., Muzquiz, J.L., Girones, O. Characterization of probiotic properties of lactic acid bacteria isolated from intestinal microbiota of fish. *Aquaculture*, 2008, 278: 188–191.
DOI: 10.1016/j.aquaculture.2008.03.014
- [28] Lazado, C.C., Caipang, C.M.A. Mucosal immunity and probiotics in fish. *Fish & Shellfish Immunology*, 2014, 39: 78–89.
DOI: 10.1016/j.fsi.2014.04.015
- [29] Farahi, A., Kasiri, M., Sudagar, M., Alamshahi, F. Angelfish (*Pterophyllum scalare* Schultze, 1823) Larvae. *Journal of Animal and Veterinary Advances*, 2011, 10: 2305-2311.
- [30] Meidong, R., Doolgindachbaporn, S., Sakai, K., Tongpim, S. Isolation and selection of lactic acid bacteria from Thai indigenous fermented foods for use as probiotics in tilapia fish *Oreochromis niloticus*. *Aquaculture, Aquarium, Conservation & Legislation-International Journal of the Bioflux Society (AACL Bioflux)*, 2017, 10: 455-463.
- [31] Rendueles, O., Ferrières, L., Frétaud, M., Bégaud, E., Herbomel, P., Levraud, J.P., Ghigo, J.M. A new zebrafish model of oro-intestinal pathogen colonization reveals a key role for adhesion in protection by probiotic bacteria. *PLoS pathogens*, 2012, 26:e1002815.
DOI: 10.1371/journal.ppat.1002815
- [32] Bledsoe, J.W., Peterson, B.C., Swanson, K.S., Small, B.C. Ontogenetic characterization of the intestinal microbiota of channel catfish through 16S rRNA gene sequencing reveals insights on temporal shifts and the influence of environmental microbes. *PloS one*, 2016, 15: e0166379.
DOI: 10.1371/journal.pone.0166379