



# Journal of Zoological Research

Volume 1 | Issue 3 | 2019 October | ISSN 2630-5100 (Online)















#### **Editor-in-Chief**

#### Dr. Guangshun Jiang

Northeast Forestry University, China

#### **Editorial Board Members**

Murat Kabak, Turkey Masoud Yousefi, Iran Anil Kumar Singh, India Nagendra Kumar Singh, India Umar Farouk Mustapha, China Nicole Fidalgo Paretsis, Brazil Antón Rafael García, Spain Sunil Kumar, India Thomas Francis Lado, South Sudan Gamaleldin Mustafa Suliman, Saudi Arabia Muammer Kurnaz, Turkey Neelam Kushwaha, India Vahid Nasiri, Iran Arunkarthick Samudram, India Somayeh Sharifi, Iran Satendra Kumar Yadav, India Mohammadreza Mohammadabadi, Iran Paolo Riccardo Martelli, Hong Kong Mariane Aparecida Nickele, Brazil José Eduardo Serrão, Brazil Halime KOÇ-GÜR, Turkey Azar Shokri, Iran Shafiya Imtiaz Rafiqi, India Zaigui Wang, China Mayra Anton Dib Saleh, Brazil Jeff M. Pérez, Qatar Arun Kumar Roy Mahato, India Gisela Ariana Marcoppido, Argentina Tirasak Pasharawipas, Thailand Hubert Untersteiner, Austria Mduduzi Ndlovu, South Africa Jan Klimaszewski. Canada Aris Pourlis. Greece Orus Ilyas, India Khaled Abouelezz Fouad Mohammed, Egypt Mohamed Abdo Rizk, Egypt Han-Tsung Wang, Taiwan Muhammad Naeem Tahir, Pakistan Arda Sözcü, Turkey Sevdan Yilmaz, Turkey Bibu John Kariyil, India Thomas Oliver Mérő, Hungary Eduardo Reséndiz, Mexico Amr Salah Morsy, Egypt Jesús Eduardo Resendiz M., Mexico Imen Belhadj-Slimen, Tunisia Masoud Yousefi, Iran Olukorede Ibukun Popoola, Nigeria Jean Fortune Dagnon, BENIN

# Journal of Zoological Research

**Editor-in-Chief** Dr. Guangshun Jiang





### Contents

## Article

1 Parasite Fauna of *Lutjanus synagris* Commercialized in the Fish Market from Bragança-PA, Brazil

Natalino da Costa Sousa Daiana Silva dos Santos Silmara Rosa Silva Alexandre Vaz da Silva Jucimauro de Araújo P. Junior João Vitor Antunes Miranda Pedro Rodrigo Nery de Souza Matheus Carvalho de Melo Fagner de Melo Xavier Wellington Lima da Silva Junior Fabricio Ramos Menezes Márcia Valeria Silva do Couto

8 Milk Fatty-Acid Profile after Feeding Increasing Doses of a Mixture of Soybean and Linseed Oils to Pasture Dairy Cows

Liliana Elisabet Antonacci Gerardo Antonio Gagliostro

23 Five years Retrospective Study of Avian Coccidiosis in a Veterinary Clinic Bukuru Plateau State Nigeria

Barde, Israel Joshua Ladan, Haruna Bello Shekaro, Audu Ijoma, Sandra Ifynneke Idachaba, Stella Ejura Olabode Victoria Bose Oguche, Moses Ojonugwa Ishaku, Bata Shalangwa

28 Contribution of Livestock Production to Global Greenhouse Gas Emission and Mitigation Strategies

Ahmedin Abdurehman Musa

## Review

5 The Importance of Motacilla Alba Behavior on Hitting Its Own Mirror Reflection Yan Zhang Greg Mirt Fan Xu Fangfang Liu

## Copyright

*Journal of Zoological Research* is licensed under a Creative Commons-Non-Commercial 4.0 International Copyright (CC BY- NC4.0). Readers shall have the right to copy and distribute articles in this journal in any form in any medium, and may also modify, convert or create on the basis of articles. In sharing and using articles in this journal, the user must indicate the author and source, and mark the changes made in articles. Copyright © BILINGUAL PUBLISHING CO. All Rights Reserved.



**Journal of Zoological Research** http://ojs.bilpublishing.com/index.php/jzr



#### ARTICLE

# Parasite Fauna of *Lutjanus synagris* Commercialized in the Fish Market from Bragança-PA, Brazil

Natalino da Costa Sousa<sup>1\*</sup> Daiana Silva dos Santos<sup>1</sup> Silmara Rosa Silva<sup>1</sup> Alexandre Vaz da Silva<sup>1</sup> Jucimauro de Araújo P. Junior<sup>1</sup> João Vitor Antunes Miranda<sup>2</sup> Pedro Rodrigo Nery de Souza<sup>2</sup> Matheus Carvalho de Melo<sup>2</sup> Fagner de Melo Xavier<sup>2</sup> Wellington Lima da Silva Junior<sup>3</sup> Fabricio Ramos Menezes<sup>4</sup> Márcia Valeria Silva do Couto<sup>1</sup>

1. Federal University of Pará (UFPA), *Campus* Bragança, Alameda Leandro Ribeiro, Bragança, 68600-000, PA, Brazil 2. Federal University of Pará (UFPA), *Campus* Tucuruí-Pará, Brazil

3. Federal University of Sergipe (UFS), Departamento de Pesca e Aquicultura. Aracaju-Sergipe, Brazil

4. Instituto Federal do Pará (IFPA), Campus de Cametá-Pará, Brazil

#### ARTICLE INFO

Article history Received: 13 April 2020 Accepted: 28 April 2020 Published Online: 30 June 2020

*Keywords:* Cymothoidae Parasite Marine fish

#### ABSTRACT

Studies about the parasite fauna of marine fish highlights as an important problem for public health with zoonotic parasites or affecting the fish quality. Thus, this study evaluated the parasite fauna of Lutjanus synagris commercialized in the fish market from Bragança-PA. In laboratory, 58 fish were measured, weighted and conducted to parasitological analysis to determine parasitological indexes and relative dominance. Every parasite was fixed and identified until to the lowest taxonomic level. Through the parasitological analysis, it found Cymothoidae, Digenea, Cucullanus sp. and Procamallanus (Spirocamallanus) sp., with total prevalence 67.24%. Digenea showed the highest prevalence and mean intensity values. For nematode, Cucullanus sp. obtained the greater prevalence and relative dominance, while Procamallanus (Spirocamallanus) sp. showed the greater mean intensity and abundance. Cymothoidae showed the lowest prevalence and mean intensity values. As conclusion, the parasite fauna of L. synagris has been noted with low diversity, reporting the nematode occurrence Procamallanus (Spirocamallanus).

#### 1. Introduction

Fishing activity, an important source of fish meat for human consumption, generated approximately 91 million of tons at 2016<sup>[1,2]</sup>. Fish species from Lutjanus family are distributed in the tropical and subtropical regions with carnivorous habit and high commercial value, widely appreciated by consumers <sup>[3,4]</sup>. Among the fish species, ariacó *Lutjanus synagris* commonly known as red fish are appreciated for all northeast region from Pará due to the meat quality and price.

Despite the commercial importance of this species for human consumption, its trade can represents risk to public health with zoonotic parasites <sup>[5,6,7]</sup>. In addition, the high

Natalino da Costa Sousa,

<sup>\*</sup>Corresponding Author:

Federal University of Pará (UFPA), Campus Bragança, Alameda Leandro Ribeiro, Bragança, 68600-000, PA, Brazil; Email: natal.engpesca@gmail.com

parasite infestation can provoke physiological alterations in the fish, making it more susceptible to diseases <sup>[8,9]</sup>.

Currently, still are few scientific papers about the parasite fauna for genus Lutjanus, with reports of crustaceans, monogenea, nematode and cestode <sup>[5,7,10,11,12]</sup>. In front of this, take knowledge about the parasite fauna can contributes with important information of national marine parasites. Thus, this study evaluated the parasite fauna of *L. synagris* commercialized in the fish market from Bragança PA.

#### 2. Material and Methods

Species of *L. synagris* were purchased between September and October at 2018 of fish market from Bragança PA (1°05'42.94" S and 47°16'19.52" W) adequately packed in plastic bags and conducted to laboratory. Every fish were identified according to Lessa and Nóbrega <sup>[13]</sup>, measured (TL: total length) and weighted (W: weight).

Parasitological analysis were carried out according to Eiras *et al.*<sup>[14]</sup> and Amato *et al.*<sup>[15]</sup>, being quantified, collected, fixed and identified until the lowest taxonomic level<sup>[16]</sup>. Based on parasite quantification, this study determined the parasitological indexes as prevalence (P), mean intensity (MI), mean abundance (MA) <sup>[17]</sup> as well as the relative dominance (RD) <sup>[18]</sup>.

#### 3. Result and Discussion

About the 67.24% of analyzed *L. synagris* (58 fish 26.55  $\pm$  5.12 cm 482.12  $\pm$  186.30 g) has been infested by parasite at least one taxon. This study identified four taxon (table 1). The scientific literature has been reported crustaceans (*Lernantrhopus* sp.) nematode (*Anisakis* sp., *Capillaria* sp. e *Cucullanus* sp.) and cestode (Larvas de *Floriceps* sp.) for *L. synagris* <sup>[5,7,10,19,20]</sup>.

 Table 1. Parasitological indexes, relative dominance and infestation site on *L. synagris* commercialized in the fish market from Bragança PA

Parasite	IS	P (%)	MI	МА	RD
Cymothoidae	G/M	10.34	1.33	0.14	0.04
Digenea		62.07	3.44	2.14	0.59
Cucullanus sp.	Ι	46.55	1.93	0.90	0.25
Procamallanus (Spirocamallanus) sp.	Ι	13.79	3.38	0.47	0.13

*Notes:* IS - Infestation site, P - prevalence, MI - mean intensity, MA - mean abundance, RD - relative dominance, G/M - gill/mouth, PS/I - pyloric cecum, I - intestine.

Digenea, found in the pyloric cecum and intestine, showed the greater prevalence, mean intensity, mean abundance and relative dominance (table 1). Within found nematode, *Cucullunus* sp. obtained the greater prevalence (46.55%) and relative dominance (0.25), while *Procamallanus* (*Spirocamallanus*) sp. showed greater mean intensity (3.38) and mean abundance (0.47).

Fish are considered intermediary host to the digenea life cycle<sup>[21]</sup>, being a common parasite into aquatic ecosystem widely reported to L. *guttatus; L. adetti* and *L. fulviflamma*<sup>[22,23,24]</sup>. According to the Argáez-García *et al.* <sup>[19]</sup>, they found digenea species *Hamacreadium mutabile*, *Helicometrina nimia*, *Metadena globosa*, *Stephanosthomum casum*, *Paracryptogonimus americanos*, *Hemiurus sp.* and *Neoprosorhynchus* in the pyloric cecum, intestine and stomach of *L. griseus*.

According to the Morales-Serna *et al.* <sup>[23]</sup>, evaluating the parasite fauna of *L. guttatus* between 2004 and 2006, they found prevalence values of 0 to 21% for digenea, lower value if compared to the present study. However, its mean intensity for the same parasite (digenea) showed greater values (4.5) than this study.

The most studies of nematode at marine fish only describes the parasite <sup>[25,26,27]</sup>. The greater importance about this parasite would be its zoonotic potential <sup>[28,29]</sup>. In the present study, found nematodes have no zootechnical potential, different result if compared to the Alves *et al.* <sup>[7]</sup> with genus *Anisakis* sp. and *Raphidascaris* sp. (Ichthyascaris) at prevalence 17.39 and 4.34% respectively for *L. synagris*. Other study on the same fish species, they found cestode larvae (*Floriceps* sp., *Pseudogrillotia* sp. *Oncomegas* sp.) with the first report about *Philometrai* sp. <sup>[20,25]</sup>, a parasite which affects the fish meat.

According to González-Solís *et al.* <sup>[30]</sup>, they identified nematode *Cucullanus* in fish species *Arothron hispidus, Abudefduf sordidus* and *Caranx ignobilis* with prevalence 47% and mean intensity (6±4.7) for *A. hispidus* species. The prevalence results were similar to the present study, this being the nematode group with greater relative dominance (0.25). None study is related to the presence of the genus Procamallanus (Spirocamallanus) in *L. synagris*, which is probably caused by the consumption of zooplankton, which is considered a parasite in the egg or larvae phase <sup>[31]</sup>.

In the present study, despite the low prevalence (10.34%), mean intensity (1.33), medium abundance (0.14) and relative dominance (0.04) observed for the crust of the Cymothoidae family, this record is relevant to survey the parasitic fauna of *L. synagris*. The reports by Cavalcante *et al.* <sup>[10]</sup> observed this same species of fish the parasites *Lernantrhopus* sp., *Lernaelophus striatus* and *Rocinela* sp. that were found in the gills and mouth. Therefore, information about a parasitological fauna of marine species with economic value in the market is essential for the management of the commercialization of this fish.

#### 4. Conclusion

The *Lutjanus synagris* has been noted with a low parasite fauna, with the greater prevalence for digenea and occurrence of nematode *Procamallanus* (*Spirocamallanus*) sp.

#### References

- [1] FAO. Food and Agriculture Organization. The State of World Fisheries and Aquaculture. Meeting the sustainable development goals. Rome, 2018: 227.
- [2] Santos, R.F., Santos, W.J.P., Monteiro, E.P., Nascimento, J. C. S. A pesca artesanal no nordeste paraense, município de Viseu-Pará. Acta of Fisheries and Aquatic Resources, 2018, 6(1): 35-42. DOI: 10.2312/Actafish.2018.6.1.35-42
- [3] Cavalcante, L.D.F.M., de Oliveira, M.R., Chellappa, S. Aspectos reprodutivos do ariacó, *Lutjanus synagris* nas águas costeiras do Rio Grande do Norte. Biota Amazônia, 2012, 2(1): 45-50.
  DOI: 10.18561/2179-5746/biotaamazonia. v2n1p45-50
- [4] Soares, D.C.E., Marques, R.R., Lima, D.S., Vale, I.B. Caracterização da pesca artesanal no município de Porto do Mangue–Rn, Brasil. Revista Brasileira de Engenharia de Pesca, 2018, 11(2): 35-43. DOI: 10.18817/repesca.v11i2.1627
- [5] Cortés, J., Valbuena, J., Manrique, G. Nemátodos parásitos de *Lutjanus synagris* (Linneaus, 1758) y Lutjanus analis (Cuvier, 1828) (Perciformes, Lutjanidae) en las zonas de Santa Marta y Neguanje, Caribe Colombiano. Revista de la Facultad de Medicina Veterinaria y de Zootecnia, 2009, 56(1): 23-31.
- [6] Fontenelle, G., Knoff, M., Felizardo, N.N., Lopes, L.M.S., São Clemente, S.C.D. Nematóides de importância zoonótica em Cynoscion guatucupa (Pisces) no estado do Rio de Janeiro. Revista Brasileira de Parasitologia Veterinária, 2013, 22(2): 281-284. DOI: 10.1590/S1984-29612013005000019
- [7] Alves, A. M., Souza, G. T. R., Takemoto, R. M., Melo, C. M., Madi, R. R., Jeraldo, V. L. S. Anisakidae Skrjabin & Karokhin, 1945 and Raphidascarididae Hartwich, 1954 nematodes in lutjanidae (pisces: perciformes) from the Brazilian Northeast Coast. Brazilian Journal of Biology, 2019, Ahead of Print.

DOI: 10.1590/1519-6984.190350

[8] Del Rio Zaragoza, O.B., Fajer Avila, E.J., Almazán Rueda, P. Haematological and gill responses to an experimental infection of dactylogyrid monogeneans on the spotted rose snapper Lutjanus guttatus (Steindachner, 1869). Aquaculture research, 2010, 41(11): 1591-1601. DOI: 10.1111/j.1365-2109.2009.02471.x

- [9] Sowjanya, P., Rajesh, K., Raju, B.P., Lakshmi, K.V., Ramulu, K.S. Histopathology of the gill of Lutjanus russelli infected with Learnanthropus species (Copepoda: Anthosomatidae). International Journal of Current Science, 2014, 10: 11-13.
- [10] Cavalcanti, E.T.S., Nascimento, W.S., Takemoto, R.M., Alves, L.C., Chellappa, S. Occurrence of ectoparasite crustaceans in ariacó fish, *Lutjanus synagris* (linnaeus, 1758), in the coastal waters of Rio Grande do Norte, Brazil. Amazon Biota, 2013, 3(1), 94-99. (in Portuguese) DOI: 10.18561/2179-5746/biotaamazonia. v3n1p94-99
- [11] Sun Y., Yang T. Two new species of Euryhaliotrema Kritsky et Boeger, 2002 (Monogenea: Dactylogyridae) from Lutjanus russellii (Bleeker) and L. argentimaculatus (Forsskål) (Teleostei: Lutjanidae) in the South China Sea. Folia parasitologica, 2015, 62: 040. DOI: 10.14411/fp.2015.040
- [12] Abdel-Baki A.A.S., Al-Qahtani H.A., Al-Quraishy S., Mansour L. Ceratomyxa azevedoi n. sp. (Myxozoa: Myxosporea) parasitizing the gallbladder of Lutjanus ehrenbergii in the Arabian Gulf. Parasitology research, 2017, 116(10): 2757-2763. DOI: 10.1007/s00436-017-5586-8
- [13] Lessa, R., Nóbrega, M.F. Marine Fish Identification Guide for Northeast Brazil. Revizee Program, Synthesis Report, Recife. 2000: 123. (in Portuguese)
- [14] Eiras J.C., Takemoto R.M., Pavanelli G.C. Study methods and laboratory techniques in fish parasitology. 2nd Ed. Eduem, Maringá. 2006: 199. (in Portuguese)
- [15] Amato J.F.R. Boeger W.A., AMATO, S.B. Laboratory protocols: Collection and processing of fish parasites. University of the Federal Rural University of Rio de Janeiro, Seropédica. 1991: 52. (in Portuguese)
- [16] Thatcher V.E. Amazon fish parasites. 2<sup>a</sup> ed. Sofia Moscow: Editora Pensoft Publishers, 2006: 118.
- [17] Bush A.O., Lafferty K.D., Lotz J.M., Shostak A.W. Parasitology meets ecology on its own terms: Margolis *et al.* revisited. The Journal of parasitology, 1997, 83(4): 575-583.
- [18] Rohde K., Hayward C., Heap M. Aspects of the ecology of metazoan ectoparasites of marine fishes. International journal for parasitology, 1995, 25(8): 945-970.

DOI: 10.1016/0020-7519(95)00015-T

[19] Argáez-García N., Guillén-Hernández S., Aguirre-Macedo M.L. Intestinal helminths of Lutjanus griseus (Perciformes: Lutjanidae) from three environments in Yucatán (Mexico), with a checklist of its parasites in the Gulf of Mexico and Caribbean region. Revista Mexicana de Biodiversidad, 2010, 81(3): 903-912.

[20] Alves A.M., Souza G.T.R., Takemoto R.M., Tavares L.E.R., Melo C.M.D., Madi R.R., Jeraldo V.D.L.S. Occurrence of larvae of trypanorhynch cestodes in snappers (Lutjanidae) from northeast Brazil. Revista Brasileira de Parasitologia Veterinária, 2018, 27(3): 415-419.

DOI: 10.1590/s1984-296120180019

- [21] Ramos I.P., Franceschini L., Zago A.C., Zica É.D.O.P., Wunderlich A.C., Carvalho E.D., Silva R.J.D. New host records and a checklist of fishes infected with Austrodiplostomum compactum (Digenea: Diplostomidae) in Brazil. Revista Brasileira de Parasitologia Veterinária, 2013, 22(4): 511-518. DOI: 10.1590/S1984-29612013000400010
- [22] Justine, J.L., Beveridge, I., Boxshall, G.A., Bray, R.A., Miller, T.L., Moravec, F., Trilles, J.P., Whittington, I.D. An annotated list of fish parasites (Isopoda, Copepoda, Monogenea, Digenea, Cestoda, Nematoda) collected from Snappers and Bream (Lutjanidae, Nemipteridae, Caesionidae) in New Caledonia confirms high parasite biodiversity on coral reef fish. Aquatic Biosystems, 2012, 8(1): 1-29. DOI: 10.1186/2046-9063-8-22
- [23] Morales-Serna, F.N., García-Vargas, F., Medina-Guerrero, R.M., Fajer-Ávila, E.J. Helminth parasite communities of spotted rose snapper Lutjanus guttatus from the Mexican Pacific. Helminthologia, 2017, 54(3): 240-249. DOI: 10.1515/helm-2017-0031
- [24] Miller, T.L., Cutmore, S.C., Cribb, T.H. Two species of Neometadena Hafeezullah & Siddiqi, 1970 (Digenea: Cryptogonimidae) from Moreton Bay, Australia, including the description of Neometadena paucispina n. sp. from Australian Lutjanidae. Systematic parasitology, 2018, 95(7): 655-664. DOI: 10.1007/s11230-018-9804-2
- [25] Cavalcanti, E.T.S., Takemoto, R.M., Alves, L.C., Chellappa, S. First record of endoparasite Philometra sp. (Nematoda: Philometridae) in lane snapper *Lutjanus synagris* from the coast of Rio Grande do Norte,

Brazil. Marine Biodiversity Records, 2010, 3: E93. DOI: 10.1017/S1755267210000862

- [26] Moravec F., Gey D., Justine J.L. Nematode parasites of four species of Carangoides (Osteichthyes: Carangidae) in New Caledonian waters, with a description of Philometra dispar n. sp. (Philometridae). Parasite, 2016, 23: 40. DOI: 10.1051/parasite/2016049
- [27] Subekti, S., Puspitarini, D. A. Identifikasi dan Prevalensi Cacing Endoparasit pada Saluran Pencernaan Kakap Merah (Lutjanus argentimaculatus) di Keramba Jaring Apung Balai Besar Perikanan Budidaya Laut, Lampung. Jurnal Ilmiah Perikanan dan Kelautan, 2018, 10(1): 59-64. Dok. 10.2172/10.1111/0.540

DOI: 10.20473/jipk.v10i1.8549

- [28] Buchmann, K., Mehrdana, F. Effects of anisakid nematodes *Anisakis* simplex (sl), Pseudoterranova decipiens (sl) and Contracaecum osculatum (sl) on fish and consumer health. Food and Waterborne Parasitology, 2016, 4: 13-22. DOI: 10.1016/j.fawpar.2016.07.003
- [29] Rodríguez, H., González, Á.F., Abollo, E., Pascual, S. Re-evaluation of anchovies (Engraulis encrasicolus) as an important risk factor for sensitization to zoonotic nematodes in Spain. Fisheries Research, 2018, 202: 49-58.

Doi: 10.1016/j.fishres.2017.11.013

[30] González-Solís, D., Soler-Jiménez, L. C., Aguirre-Macedo, M. L., McLaughlin, J. P., Shaw, J. C., James, A. K., Hechinger, R.F., Kuris, A.M., Lafferty, K.D., Vidal-Martínez, V. M. (2019). Parasitic nematodes of marine fishes from Palmyra Atoll, East Indo-Pacific, including a new species of Spinitectus (Nematoda, Cystidicolidae). ZooKeys, 2019, 892: 1-26.

DOI: 10.3897/zookeys.892.38447

[31] Fujimoto, R.Y., Couto, M.V.S., Sousa, N.C., Riscala, R., Eiras, J.C., Laterça, M. Seasonality of infection by *Procamallanus (Spirocamallanus)* inopinatus (nematoda: camallanidae) in Bryconops melanurus (characiformes: iguanodectidae). Fisheries Institute Bulletin, 2018, 44(4): 331-338. DOI: 10.20950/1678-2305.2018.44.4.334



Journal of Zoological Research

http://ojs.bilpublishing.com/index.php/jzr



# **REVIEW** The Importance of Motacilla Alba Behavior on Hitting Its Own Mirror Reflection

### Yan Zhang<sup>1</sup> Greg Mirt<sup>2</sup> Fan Xu<sup>1\*</sup> Fangfang Liu<sup>3\*</sup>

1. Department of Public Health, Chengdu Medical College, Sichuan, 610500, China

2. Occupational Activity Centre Novo mesto, Slovenia, EU

3. Art college, Southwest Minzu University, Sichuan, 610041, China

ARTICLE INFO	ABSTRACT
Article history Received: 10 June 2020 Accepted: 17 June 2020 Published Online: 30 June 2020	Self-awareness is considered as a capability of recognize oneself and increasingly received attention. However, self-awareness in the bird Motacilla Alba is unclear. To study the self-recognition in Motacilla Alba, the subject is observed by mirror while eating. The bird performed the look around, confirm again the surroundings, become alert, hit the mirror.
Keywords:	These behaviors suggests that presently Motacilla Alba does not have the capacity of self-awareness by the test.
Motacilla Alba	
Self-awareness	
Mirror Self-Recognition (MSR)	

#### 1. Self-awareness Process

The ability to recognize oneself in the mirror is considered as self-awareness<sup>[1]</sup>. Mirror self-recognition (MSR) is designed as a method to explore the animal's sense of self<sup>[2]</sup>. Tests of mirror self-recognition (MSR) have been central to our understanding of self-awareness from developmental and evolutionary perspectives.

Self-awareness in humans spontaneously emerged <sup>[3]</sup>. In 2000,Kusayama et al showed that four jungle crows (Corvus macrorhynchos) were exposed to a mirror with peck and flap behaviors but species failed to pass the MSR <sup>[4]</sup>. In 2002, Watanabe experimented that java sparrows were equaled to choose a mirror and a frosted live bird when they were exposed to a mirror and a frosted mirror. The

\*Corresponding Author:

result suggested that java sparrows saw the self-image on the mirror as conspecific image <sup>[5]</sup>. Furthermore, In 2017, Fanny-Linn Kraft tested the great tit mirror response with social behavior. The result showed no evidence that the great tit possessed self-awareness <sup>[6]</sup>.

#### 2. Fly to Mirror Behavior



Figure 1. Check the subject in the mirror

Fangfang Liu,

Art college, Southwest Minzu University, Sichuan, 610041, China; Email: 619898782@qq.com

Fan Xu,

Department of Public Health, Chengdu Medical College, Sichuan, 610500, China; Email: xufan@cmc.edu.cn

#### 3. Meanings



Figure 2. Check who/which hide behind the mirror



Figure 3. Get angry about the unknown subject



Figure 4. Fly to Mirror Behavior

Interestingly, we have set up a mirror on the ground in the open field. The bread was placed in front of the mirror. In this test, we found that Motacilla alba looked around before it approached the bread, see Figure 1. The reflection of mirror was found by the subject while picking food. It became more nervous. Then it walked to the back of the mirror to confirm whether there is conspecifics behind of the mirror for three times, see Figure 2, 3. At the beginning, the subject picked the food peacefully. It became more and more alert when it saw the reflection in the mirror. Ultimately, Motacilla alba violently attacked the reflection continuously in the mirror and a bright chirp, see Figure 4.

The MSR is an excellent method to test self-awareness. Motacilla alba presented the aggressive behavior when saw its reflection in the mirror. Unfortunately, neither of these behaviors could be demonstrate that the Motacilla alba can recognized itself in the mirror.



Figure 5. Aggressive behavior-Fly to mirror

So far, most of the species on the MSR tested have failed to identify itself in the mirror. Many of them responded to their self-image with social behavior if considered the image their conspecifics. Some species have shown aggressive behavior <sup>[7]</sup>. Even humans aren't born with a sense of self. We may not recognize ourselves in a mirror from 15 to 24 months of age <sup>[8]</sup>. There is an important evidence that only a few species can pass the MSR test, and only long-term use of mirrors as visual stimuli can pass the MSR test such as (four great apes<sup>[9]</sup>, bottlenose dolphins <sup>[10]</sup>. Asian elephants <sup>[11]</sup>, and magpies <sup>[12]</sup>. Some species have the possibility to modify their self-characteristics through learning <sup>[13]</sup>. This proves to a certain extent that self-consciousness requires certain conditions, which can be learned. However it cannot be absolutely denied that those animals (fishes, birds, sea lions, dogs and cats <sup>[14]</sup>) have no sense of self. It only proves that they are not self-aware at this stage. When the birds are facing the similar selection pressures that leads to changes in neural structure, particularly in cognition<sup>[15]</sup>. Our data demonstrated that Motacilla alba are not capable to recognize itself in the mirror for the first time.

#### Funding

This work was supported by the Chengdu Medical College Natural Science Foundation (CYZ18-33).

#### References

- Huttunen A. W., et al. Can self-awareness be taught? Monkeys pass the mirror test-again. Proc Natl Acad Sci U S A, 2017, 114(13): 3281-3283.
- [2] Gallop G. G., Jr. Chimpanzees: self-recognition. Science, 1970, 167(3914): 86-87.
- [3] Morrison R., D. Reiss. Precocious development of self-awareness in dolphins. PLoS One, 2018, 13(1):

e0189813.

- [4] Kusayama T., et al. Responses to mirror-image stimulation in jungle crows (Corvus macrorhynchos). 2000, 3(1): 61-64.
- [5] Watanabe, S. Preference for mirror images and video image in Java sparrows (Padda oryzivora). Behavioural processes, 2002, 60: 35-39.
- [6] Kraft F. L., et al. No evidence for self-recognition in a small passerine, the great tit (Parus major) judged from the mark/mirror test. Anim Cogn, 2017, 20(6): 1049-1057.
- [7] Deregnaucourt, S., D. Bovet. The perception of self in birds. Neurosci Biobehav Rev., 2016, 69: 1-14.
- [8] Amsterdam B. Mirror self-image reactions before age two. Dev Psychobiol, 1972, 5(4): 297-305.
- [9] Anderson J. R., Gallup, G. G. Which Primates Recognize Themselves in Mirrors? PLoS Biology, 2011, 9(3): e1001024.
- [10] Reiss, D., Marino, L. Mirror self-recognition in the bottlenose dolphin: A case of cognitive convergence.

Proceedings of the National Academy of Sciences, 2001, 98(10): 5937–5942.

- [11] Plotnik, J. M., de Waal, F. B. M., Reiss, D. Self-recognition in an Asian elephant. Proceedings of the National Academy of Sciences, 2006, 103(45): 17053–17057.
- [12] Prior, H., Schwarz, A., Güntürkün, O. Mirror-Induced Behavior in the Magpie (Pica pica): Evidence of Self-Recognition. PLoS Biology, 2008, 6(8): e202.
- [13] Bolhuis, J. J., Okanoya, K., Scharff, C.. Twitter evolution: converging mechanisms in birdsong and human speech. Nature Reviews Neuroscience, 2010, 11(11): 747–759.
- [14] Parker, S., Mitchell, R., Boccia, M. (Eds.). Self-Awareness in Animals and Humans: Developmental Perspectives. Cambridge: Cambridge University Press, 1994.
- [15] Edelman, D. B., Seth, A. K. Animal consciousness: a synthetic approach. Trends in Neurosciences, 2009, 32(9): 476–484.



**Journal of Zoological Research** http://ojs.bilpublishing.com/index.php/jzr



# ARTICLE Milk Fatty-Acid Profile after Feeding Increasing Doses of a Mixture of Soybean and Linseed Oils to Pasture Dairy Cows

#### Liliana Elisabet Antonacci Gerardo Antonio Gagliostro<sup>\*</sup>

Instituto Nacional de Tecnología Agropecuaria (INTA), Estación Experimental Balcarce, Área de Producción Animal, Balcarce, Argentina

ARTICLE INFO	ABSTRACT
Article history Received: 12 June 2020 Accepted: 27 June 2020 Published Online: 30 June 2020	The goal was to determine the effect of growing intake of a mixture (75:25) of soybean (SoOi) and linseed (LiOi) oils on milk production and composition and milk fatty-acid (MF-A) profile in grazing dairy cows. Twenty-four Holstein cows were assigned to 4 treatments in a completely randomized design with three weeks of adaptation to oil doses and one week of experi-
Keywords: Polyunsaturated oils Vegetable oils Rumenic acid	mental measurements. On a dry matter (DM) basis, cows were fed pasture (63%), energy concentrate (37%) and the SoOi LiOi oil mixture at zero (Tr0%), 2% (Tr2%), 4% (Tr4%) and 6% (Tr6%) of total DM intake equivalent to 0, 0.36, 0.72 and 1.08 kg/cow/day of the oil mixture. The oil mixture was manually mixed-up to the concentrate (7.04 kg DM/cow/day) and supplied by halves during each milking time without refusals. Pasture ( $P = 0.49$ ) and total DM intakes ( $P = 0.31$ ) were similar between treatments averaging 11.27 and 18.85 kg DM/cow/day respectively. Milk output (22.71 kg/cow/day) was not affected ( $P = 0.46$ ). Milk fat content reduced linearly ( $P < 0.05$ ) from 3.20 (Tr0%) to 2.67 g/100g (Tr6%) without effects ( $P = 0.73$ ) on fat or fat corrected milk (4%FCM) yields. Milk protein concentration ( $P < 0.56$ ) or yields ( $P < 0.11$ ) were not affected. Lactose contents tended ( $P < 0.08$ ) to be higher in oil supplemented cows and milk urea nitrogen was not affected ( $P = 0.14$ ). The basal (Tr0%) concentration (g/100g MF-A) of totaly hypercholesterolemic MF-A ( $C_{12.0}$ , $C_{14.0}$ and $C_{16.0}$ ) averaged 38.93 and decreased linearly ( $P < 0.0001$ ) with oil intake to 37.81 (Tr2%), 31.59 (Tr4%) and 29.18 (Tr6%). Levels of elaidic ( <i>trans</i> -9 $C_{18.1}$ ) and <i>trans</i> -10 $C_{18.1}$ MF-A resulted low-slung in the basal (Tr0%) milk (0.21 and 0.20 g/100g MF-A, respectively) but increased linearly ( $P < 0.0001$ ) after oil intake reaching the maximum values at Tr6% (0.73 and 2.23 g/100g MF-A, respectively). Milk concentration (g/100g MF-A) of vaccenic acid ( <i>trans</i> -11 $C_{18.1}$ , VA) averaged 3.63 in Tr0% and increased linearly ( $P < 0.0001$ ) with oil intake reaching 4.97, 7.05 and 8.38 in Tr2%, Tr4% and Tr6%, respectively. Basal concentration of rumenic acid ( <i>cis</i> -9. <i>trans</i> -11 $C_{18.2}$ , RA) was 2.28 g/100g MF-A, and increased linearly ( $P < 0.0001$ ) with increased oil dose resulting in maximal plateau in Tr4% (3.88) and Tr6% (3.89). The basal atherogenic index (AI) of milk was 1.87 and linearly decreased ( $P < 0.01$ ) t
*Corresponding Author:	ratio with low levels of the detrimental trans-9 $C_{18:1}$ and trans-10 $C_{18:1}$ .
Gerardo Antonio Gagliostro, Instituto Nacional da Tannología Aguaracuaria	(INTA) Estación Experimental Palaguas Área de Dreducción Animal Palaguas
Instituto Nacional ae Iecnologia Agropecuaria	(IIVIA), Estacion Experimental Balcarce, Area de Produccion Animal, Balcarce,

Instituto Nacional de Tecnología Agropecuaria (INTA), Estación Experimental Balcarce, Área de Producción Animal, Balcarce, Argentina;

Email: gagliostro.gerardo@inta.gob.ar

#### 1. Introduction

ilk F-A composition is a determinant factor of its healthy properties due to the potential effects that certain specific MF-A have on human health. In human diets, dairy fat can account up to 75% of total consumption of fat from ruminant origin and although dairy products (which have a very low cholesterol content) provide only 15-25% of the total fat, they provide about 25 to 35% of total milk saturated fat (MSF) consumed daily<sup>[1]</sup>.

Some saturated FA present in milk just like lauric ( $C_{12:0}$ ). myristic ( $C_{14:0}$ ) and palmitic ( $C_{16:0}$ ) are potentially atherogenic when consumed in excess <sup>[2,3]</sup> and related with the increased risk of cardiovascular diseases <sup>[2,4]</sup>. Feeding oils high in polyunsaturated PUFA is an effective and natural tool to inhibit *de novo* mammary synthesis of milk saturated FA (MSF-A) reducing the presence of the pro-atherogenic MF-A of milk fat <sup>[5,6]</sup>.

A special interest has been placed in RA, the *cis*-9. *trans*-11C<sub>18:2</sub> isomer of conjugated linoleic acid (CLA), for its potential healthy role on the levels and composition of circulating lipids, cardiovascular health <sup>[7,8]</sup> and the reduction in the incidence of some types of cancer <sup>[4,9,12]</sup> and immune response <sup>[13,14]</sup>. On the other hand, VA (*trans*-11 C<sub>18:1</sub>) is the main trans MF-A being the most important precursor of RA <sup>[1]</sup>. It showed antiproliferative properties itself or after being converted to RA in human tissues at an estimated rate of 20% <sup>[15]</sup>.

Dairy fat is the most important natural source of RA and its concentration in milk fat is highly dependent on type of diet and lipid supplementation <sup>[1,5,6,16]</sup>. A pasture-based diet allows to obtain a milk with a high basal level of RA which can be amplified feeding vegetable oils high in PUFA<sup>[1,5,6,17]</sup>. A mixture (75:25) of SoOi and LiOi showed to be very effective <sup>[6]</sup> but the optimal level of oilblend supply has not yet been well defined. This is a subject of concern taking into account oils costs, deviations towards unhealthy trans-MF-A (trans-9 and trans-10 C<sub>18:1</sub>) synthesis owing to oil overdoses and the potential deleterious effects of free oils on ruminal function and digestion. Despite the practical importance of knowing what is the most adequate quantity of oils to be supplied, in our knowledge experimental results are still very scarce or directly non-existent. A linear effect of oil intake on milk RA was postulated reaching a plateau at an oil dose of 4% of total DM intake <sup>[1]</sup>. The objective of the study was to define the most adequate quantity of the SoOi LiOi mixture (75:25) to be supplied to grazing dairy cows in order to obtain milk with up high level of CLA and reduced concentrations of unhealthy fatty acids at the lowest cost.

#### 2. Materials and Methods

#### 2.1 Treatments, Animals and Experimental Design

The experiment was carried out at the National Institute of Agricultural Technology (INTA) in Balcarce (37°45'S, 58°18'W) during September and October of 2014. The experimental period lasted 4 weeks (wk) with the first three wk as adaptation to oils intake and the fourth wk for data collection. Twenty-four multiparous Holstein cows (552  $\pm$  50 kg, body weight, BW) in mid lactation (244  $\pm$ 69 days postpartum) and producing  $20.5 \pm 1.8$  kg milk/ day were allocated to four treatments (6 cows per treatment) in a completely randomized design. Treatments were defined by the increasing intake of a blend (w/w) of 75% (SoOi) and 25% (LiOi). The oil blend was consumed at 0% (Tr0%), 2% (Tr2%), 4% (Tr4%) and 6% (Tr6%) of total DM intake of the dairy cow (18 kg DM/cow/day) measured during the first wk prior to the start of the trial. Procedures and animal care were approved by the Institutional Committee for the Care and Use of Experimental Animals (CICUAE, INTA CERBAS).

A perennial pasture of brome grasse (Bromus unioloides) and red clover (Trifolium pratense) was offered using a daily-strip grazing system. The area of each strip was regulated using a temporary electric fence to provide an herbage allowance (HA) of 27 kg DM/cow/day. The available total biomass (kg DM/ha) was estimated every week to adjust the size of the daily grazing strip by the double sampling method using the relationship between the height of the forage (x) and the available biomass (y) as described previously<sup>[18]</sup>. The equations were adjusted for both, initial availability and for the remaining forage after grazing. The concentrate included ground corn grain (35%), malt brewery waste (10%), pelletized sunflower meal (20%), soybean grains (10%), wheatgrass (21.48%), calcium carbonate (2%), magnesium oxide (0.4%), salt (1%), rumensin (0.02%) and a vitamin-mineral mix (0.1%). It was offered at a rate of 8 Kg/cow/day in two equal feedings during each milking time (06.00 and 16.00).

According to treatments, the daily dose of the oil-blend was manually mixed to the concentrate during each milking time and thoroughly consumed by cows. The effective quantities of the oil-blend consumed (kg/cow/day) were 0.36 (Tr2%), 0.72 (Tr4%) and 1.08 kg (Tr6%). Adaptation to oil intake proceeded gradually by feeding by halves the target daily dose at each milking time starting with 0.1 (Tr2%), 0.2 (Tr4%) and 0.3 (Tr6%) Kg/cow/day during the first day, 0.2, 0.3 and 0.4 Kg/cow/day for the next 2 days, 0.2, 0.3 and 0.5 Kg/cow/day at day 4 and full dose according to treatment from day 5 until the end of the trial.

The animals were milked twice a day at 6:00 a.m. and 4:00 p.m. and after each milking they were conduced to the pasture with fresh and clean water available *ad libitum*.

#### 2.2 Sampling Measurements and Laboratory Procedures

Representative samples (0.5 kg) of pasture and concentrate were taken weekly. Pasture samples were collected from the grazing horizon by hand-plucking <sup>[19]</sup>. All samples were dried at 60°C for 48 hours in an oven with forced air circulation to determine DM content and then milled in a Willey mill (1 mm mesh). They were assayed for organic matter (OM) (muffle at 550-600°C for 4 hours), crude protein (CP)<sup>[20]</sup> with a LECO FP-528 analyzer), water soluble carbohydrates (WSC)<sup>[21]</sup>, neutral (NDF) and acid (ADF) detergent fiber (using the filter bag technique<sup>[22]</sup> and <sup>[23]</sup> respectively) with an autoanalyzer (ANKOM Corp. Fairtport. New York. USA 1970). Ether extract (EE) was determined by the solvent extraction technique <sup>[24]</sup> with an autoanalyzer (ANKOM Corp. Fairtport. New York. USA). The in vitro DM digestibility (IVDMD) was estimated after 48 hours of incubation in a Daisy II ANKOM equipment. Starch content was determined as described in <sup>[25]</sup>.

Pasture DM intake was individually estimated by the difference method <sup>[19]</sup> during 3 consecutive days in the week prior to the start of the experiment and during the last 3 days of week 4<sup>th</sup>. The average DM intake of the three consecutive days of measurements from each cow was computed for the statistical analysis.

Milk production was daily recorded over the whole experiment. Milk samples (50 ml) were collected at a.m. and p.m. milkings twice a week on non-consecutive days. The two samples were pooled according to the corresponding volume measured at each milking time and analyzed for fat, total protein, lactose, total and not-fat solids by mid-infrared spectrophotometry (Milko Scan-Minor, Foss Electric, Hillerod, Denmark). Milk urea nitrogen (MUN) was determined using a commercial enzymatic kit (Wiener Laboratories., Rosario, Argentina).

The cows were weighed on two consecutive days after the morning milking on days 6<sup>th</sup> 7<sup>th</sup>, 27<sup>th</sup> and 28<sup>th</sup> of the trial. Body weight (BW) change was calculated as the difference between the final minus the initial BW (average of two days) divided by the number of days elapsed. During the last two weeks of the trial, blood samples were taken by jugular vein puncture after the a.m. milking. Blood was collected in tubes containing EDTA (7-8 drops/tube, 0.342 mol/l, pH 7.2, Wiener Laboratory, Rosario. Argentina) and centrifuged ( $2000 \times g$  for 15 min at 4°C). Plasma was collected and stored at -24°C until analysis for glucose, plasma urea nitrogen (PUN), triglycerides, and cholesterol using enzymatic kits (Wiener Laboratories, Rosario, Argentina). Non-esterified fatty acids (NEFA) were assayed using the enzymatic kit from Randox Laboratories Ltd (UK).

At day 21<sup>st</sup> of oil-blend supplementation and from each composite sample collected to determine the chemical composition of milk, aliquots of 50 ml were frozen (-24°C) to obtain a single pool sample per cow for the determination of MF-A composition by gas liquid chromatography (GLC) as previously described <sup>[6]</sup>. Total milk fat (TMF) was determined gravimetrically by extraction with petroleum ether at 65-80°C<sup>[26]</sup>. The lipids were extracted with a mixture of hexane: isopropanol (3: 2) and 6% sodium sulfate at room temperature <sup>[27]</sup>. The lipid residue was dried at 40°C under a stream of nitrogen. For FA methyl esters (FAME) preparation, a cold method with hexane and 2N KOH in methanol was used <sup>[28]</sup>. The FAME were quantified using a gas chromatograph (GLC-Shimatzu GC-2014. Shimadzu Corporation. Kyoto. Japan) equipped with a CP-Sil 88 capillary column (100 mx 0.25 mm id., Varian. Lake Forrest. CA. USA) and a flame ionization detector. Injector and detector temperatures were maintained at 250°C, the flow rate at 1:100 and 1 µl of standard or milk sample using an automatic sampling device at each run of the GLC <sup>[29]</sup>. The hydrogen flow was fixed at 1 ml/min and the nitrogen flow (compression gas) at 25 ml/min. Maximum retention times and area percentages of total FA were identified by injecting known patterns. Internal standards [(Tritridecanoin [13: 0-triacylglycerol (TAG)], external reference standards GLC-463 (mixtures of 52 EMAG (purity> 99%) and trans-mix GLC 481 (purity> 99%) were purchased from Nu-Chek (Nu-Chek Prep. Inc., Elysian. MN. USA). Methyl esters of linoleic acid, cis/trans mixture (Catalog No. 47791), mixtures of unsaturated C<sub>4</sub>-C<sub>24</sub> chain length methyl esters (Catalog N° 18919) and of the individual chain length FAMEs from C4:0 to C24:1 saturated and unsaturated were obtained from Supelco (Bellefonte. PA. USA). Mixtures of positional and geometric FA isomers were provided by the CYT-ED International Network (208RTR0343). The FAME were identified by comparing their retention times with commercial standards. The values were expressed as a percentage of the total FAME. The lower limit of quantification for the FAMEs identified varied from 0.01% to 0.03%. To convert g FAME/100g methys esthers to equivalents of triacilglycerides (TAG) (g of FA as TAG/100g of total TAG) the respective Conversion Factors tabulated in the AOCS Method Ce 1j-07<sup>[29]</sup> were used. To estimate the g of MF-A 100g of sample, the g of MF-A 100g of TAG were multiplied by the total fat content (%). The results are expressed in g/100g of total MF-A.

#### 2.3 Statistical Analyses

The effect of increased levels of the oil-blend intake on milk production and composition, MF-A profile and BW changes was analyzed by orthogonal contrasts taking into account the linear, quadratic and cubic effects using the PROCEDURE MIXED<sup>[30]</sup>. Results of DM intake and plasma metabolites were analyzed by PROCEDURE GLM<sup>[30]</sup> using the following model:

$$Y_{ij} = \mu + T_i + E_{(i)}$$

Where:  $Y_{ij}$  = the dependent variable,  $\mu$ : overall mean,  $T_i$  = treatment effects and E(ij) = the residual error associated with the *ij* observation. The threshold of statistical significance was stated at P < 0.05.

#### 3. Results and Discussion

#### 3.1 Forage and Oil Charasteristics

In the pre-grazing strips, pasture biomass averaged 2100  $(\pm 308)$  kg DM/ha being above the critical value of 2000 kg DM/ha below which DM intake could be restricted <sup>[31]</sup>. The daily-strip grazing system used allowed to maintain the target HA of 27 (± 2) kg DM/cow/day adequate to obtain *ad libitum* pasture intake <sup>[31]</sup>. It has also been postulated that the maximum pasture DM intake would be achieved when HA ranges from 45 to 55 g DM/ kg BW/day<sup>[32]</sup>. For the average BW of cows (554 kg, Table 6), the HA range would be 25 to 30 kg DM/cow/day and therefore the 27 kg DM/cow/day obtained fell inside the proposed range. Pasture intake may also be affected if forage DM content is less than 18% following the linear relationship observed between pasture DM content and intake in the range of 13-22% DM content <sup>[33]</sup>. In our trial, pasture DM content averaged 20.5% (Table 1) therefore exceeding the critical range reported <sup>[33]</sup>. In the same way, pasture CP (24.1%) and NDF (36.4%) contents (Table 1) were found within the range of 15-25% (CP) and 36-54% (NDF) proposed by <sup>[31]</sup> to obtain a high forage digestibility as observed (70.93%) in vitro (Table 1). The WSC and EE contents may be considered normal for good quality pastures. It can be concluded that both, the quality and the amount of the pasture offered to cows were sufficient to achieve adequate DM and energy intakes. The chemical composition of the concéntrate (Table 1) was normal for a good quality energy concentrate.

Table 1. Chemical	composition	and in vitro	dry matter
(DM) digestibil	ity of pasture	e and concer	itrate <sup>(1)</sup>

Pasture <sup>2</sup>	Concentrate
$20.50\pm0.70$	$89.6\pm0.65$
$91.66\pm0.55$	$92.80\pm0.46$
$24.10 \pm 1.56$	$17.32\pm1.02$
$36.40 \pm 1.00$	$23.97\pm2.00$
$18.23 \pm 1.11$	$11.51 \pm 1.41$
$70.93\pm0.40$	$75.14 \pm 1.88$
$1.57 \pm 0.40$	$32.59 \pm 4.05$
$3.23\pm0.31$	$4.47\pm0.77$
$2.89\pm0.01$	$2.71\pm0.07$
12.00± 3.40	20.80± 1.51
	Pasture <sup>2</sup> $20.50 \pm 0.70$ $91.66 \pm 0.55$ $24.10 \pm 1.56$ $36.40 \pm 1.00$ $18.23 \pm 1.11$ $70.93 \pm 0.40$ $1.57 \pm 0.40$ $3.23 \pm 0.31$ $2.89 \pm 0.01$ $12.00 \pm 3.40$

*Note:* <sup>1</sup> Values expressed as the mean  $\pm$  standard deviation. Pasture and concentrate (n = 4). <sup>2</sup> Consociated pasture containing Bromus unioloides and Trifolium pratense.

The MF-A composition of feedstuffs and oils used in the experiment is shown in Table 2.

Table 2. Fatty acids composition of feedstuffs and oils

Fatty agid	Pasture <sup>1</sup>	SoOi <sup>2</sup>	LiOi <sup>3</sup>	Concentrate				
ratty actu	g/100g FA							
C <sub>16:0</sub>	14.08	10.67	6.71	13.91				
C <sub>18:0</sub>	1.31	4.31	5.38	1.92				
<i>cis-9</i> C <sub>18:1</sub>	1.53	17.43	18.28	26.89				
cis-11 C <sub>18:1</sub>	0.18	2.01	3.16	4.13				
cis-9 cis-12 C <sub>18:2</sub>	14.58	52.87	16.35	49.75				
cis-9 cis-12 cis-15 C18:3	62.76	11.36	49.87	2.29				

*Note:* <sup>1</sup>Consociated pasture containing Bromus unioloides and Trifolium pratense. <sup>2</sup>Soybean oil. <sup>3</sup>Linseed oil.

As expected, pasture and LiOi were rich in linolenic acid (*cis-9 cis-12 cis-15* C<sub>18:3</sub>). The average linoleic acid content of the pasture resulted higher than reported in other experiments <sup>[34,35]</sup> probably due to the high quality of the pasture used in the present trial. The SoOi was characterized by its high linoleic acid (*cis-9 cis-12* C<sub>18:2</sub>) content (52.87%) and by a low SFA concentration. Concentrate and oils were a good source of oleic acid (*cis-9* C<sub>18:1</sub>) as reported previously <sup>[36,39]</sup>.

When a high quality pasture is included in cow's diet, rumen lipid metabolism is oriented to healthy changes in MF-A composition mainly concerning PUFA of the  $\Omega$ 3 series and CLA. A significant reduction of MSF-A-content and the increase of oleic acid are also expected <sup>[40]</sup>. Intake of high quality fresh pastures also prevents the shift and increase in concentration of unhealthy MF-A like *trans*-9 and *trans*-10 C<sub>18:1</sub> isomers.

#### **3.2 Dry Matter Intake, Milk Yield and Composi**tion

Feeding increased doses of the oil-blend mixed to the con-

centrate did not affect (P > 0.05) concentrate, pasture or total DM intake of cows (Table 3).

**Table 3.** Intake of pasture, concentrate and oil in dairy cows supplemented or not (Control) with a blend of soybean (75%) and linseed (25%) oils at 2%, 4% and 6% of total DM intake

Parameter	Parameter Treatment <sup>1</sup>						
Intake, kg/DM/cow	Control	Tr2%	Tr4%	Tr6%	SEM	r - value	
Pasture <sup>3</sup>	12.00	10.83	10.67	11.57	0.59	0.49	
Concentrate	7.04	7.04	7.04	7.04	-	-	
Oil-blend	0.00	0.36	0.72	1.08	-	-	
Total DM	19.04	18.23	18.43	19.69	0.68	0.31	

*Note:* <sup>1.</sup> Values are expressed as LS Means and standard error of least squares means (SEM). <sup>2.</sup> Treatment effect. <sup>3.</sup> Consociated pasture containing Bromus unioloides and Trifolium pratense.

Feeding the free oil-blend could adversely affect ruminal NDF digestion <sup>[41]</sup> and reduces DM intake <sup>[42]</sup> and milk production <sup>[43,45]</sup>. Effects of free oil feeding on ruminal digestion are variable, including negative <sup>[42]</sup>, neutral <sup>[16,46,49]</sup> or even positive effects <sup>[50,51]</sup>. Inclusion of LiOi at 3.2% ( $\pm$  1.7) or SoOi at 2.9% ( $\pm$  1.2) of total DM did not affect DM intake <sup>[34,36,37]</sup>. The forage concentrate ratio (F:C) seems to interact with effects of free oil supplementation on ruminal digestion <sup>[52]</sup>. When LiOi was included at 3% of DM in a F:C ratio of 65:35, positive effects on NDF digestion were observed with an opposite result when the F:C ratio was 35:65 <sup>[51]</sup>. In the present trial, the F:C ratio averaged 61:39 (Table 3) and DM intake was not affected.

Estimated intake of linoleic and linolenic acids from the oil-blend was 149.7 and 71.9 g/cow/day in Tr2%, 299.4 and 143.8 g/cow/day in Tr4% and 449.1 and 215.7 g/cow/day in Tr6%, respectively. As negative effects on total DM intake were not observed (Table 3), energy intake would have been higher in oil-supplemented cows but yields of milk or 4%FCM remained unchanged (Table 4).

**Table 4.** Milk production and composition from grazing dairy cows supplemented or not (Control) with a blend of soybean (75%) and linseed (25%) oils at 2%, 4% and 6% of DM intake.

Parameter		Treatn	nent <sup>1</sup>			P-value		
	Control	Tr2%	Tr4%	Tr6%	SEM	Treat <sup>2</sup>	Lin <sup>3</sup>	Quad <sup>3</sup>
Milk, kg/cow/day	21.96	20.58	23.99	24.30	1.90	0.46	NS	NS
4%FCM4, kg/cow/ day	19.32	17.93	20.61	19.45	1.62	0.73	NS	NS
Fat, kg/cow/day	0.706	0.641	0.724	0.652	0.06	0.73	NS	NS
Fat, g/100g	3.20 <sup>a</sup>	3.14 <sup>a</sup>	3.06 <sup>ab</sup>	2.67 <sup>b</sup>	0.14	0.06	0.05	NS
Protein, kg/cow/ day	0.831	0.774	0.919	0.963	0.06	0.11	NS	NS
Protein, g/100g	3.81	3.80	3.87	3.99	0.11	0.56	NS	NS
Lactose, g/100g	4.80	4.85	4.93	5.06	0.07	0.08	NS	NS
Total solids, g/100g	12.74	12.85	12.82	12.61	0.23	0.85	NS	NS
Urea, mg/dl	37.83	37.11	33.30	33.12	1.76	0.14	NS	NS

*Note:* <sup>1</sup>Values expressed as least squares means and standard error of least squares means (SEM). <sup>2</sup>Treatment effect. <sup>3</sup>Contrasts: linear (Lin) and quadratic (Quad). a,b Means in the same row with different superscripts differ significantly (P < 0.05). NS = Not significant effect.

Supplementation at 4% of total DM intake with SoOi or LiOi alone or in combination (50-50) increased milk production (+16.7%) without differences between both oils <sup>[36]</sup>. In our previous trial, the average increase in milk production after oil feeding over unsupplemented cows was moderate (9.4%) and mainly explained by SoOi-LiOi mixtures at a ratio of 75-25<sup>[6]</sup>. In non-grazing trials, a high frequency of favorable effects on milk production after the inclusion of unprotected vegetable oils in the diet was reported <sup>[53]</sup>. Feeding LiOi at 3 or 4% of DM increased milk yield [30] a result that was no observed in other experiments <sup>[34,54]</sup>. When SoOi was fed at 2.9 ( $\pm$  1.3)% of the DM ration (0.533  $\pm$  0.228 kg/ cow/day) milk yield was not affected in the experiments reviewed by <sup>[37]</sup> and also when SoOi was supplied at 3.5 to 5% of total DM intake [55,57]. Supplementation with SoOi (1 to 7% of total DM) did not affect milk production [34,35,37,55]

Milk fat content decreased linearly (P < 0.05) as intake of the oil-blend increased (Table 4) an effect mainly explained by the significant decrease (-13%) observed in Tr6%. The significant reduction in the concentration of *de novo* synthesized MF-A (-100g/kg) in Tr6% compared to Control (Table 7) was not apparently compensated by a concomitant increase in mammary uptake of the preformed MF-A (+137 g/kg) since milk fat content was lower (Table 4). The result was in turn consistent with the highest concentration of the *trans*-10 C<sub>18:1</sub> isomer in milk (Table 7) since both parameters correlated negatively (Figure 1).

A direct relationship between increasing levels of *trans*-10  $C_{18:1}$  in milk and the reduction of *de novo* MF-A mammary synthesis has been reported <sup>[58]</sup> which contributes to explain the linear drop in milk fat content (Table 4). The observed fall in milk fat content was explained in part by the lower presence of the hypercholesterolemic MF-A (Table 7), which improves the healthy value of milk.



**Figure 1.** Relationship between milk fat content and *trans*-10  $C_{18:1}$  in milk from cows supplemented or not (Control) with a blend of soybean (75%) and linseed (25%) oils at 2%, 4% and 6% of DM intake

The decrease in milk fat content after PUFA oil intake is a well-documented result <sup>[36,55,57,59,60]</sup>. In an extended range (0.2 to 1.0 kg/cow/day) of unsaturated lipid supplementation to grazing dairy cows, an average decrease of of 8% in milk fat concentration and secretion has been reported <sup>[17]</sup>. Mammary uptake of certain preformed FA (*trans*-10 C<sub>18:1</sub>, *trans*-10, *cis*-12 CLA and *trans*-8, *cis*-10 CLA) reduces the activity and/or expression of genes encoding important enzymes involved in the capture, synthesis and desaturation of MF-A <sup>[61]</sup> contributing to explain the reduction in milk fat content.

Yield of 4%FCM was not different between treatments (Table 4) suggesting that the numerical increase in milk production in Tr4% and Tr6% compensated for the reduction in milk fat content. These results were consistent with the effects of unsaturated lipid supplementation that generally shows neutral effects on 4%FCM yield in confined <sup>[62]</sup> and in pasture-based diets <sup>[17]</sup>.

The lack of negative effects on milk protein concentration (Table 4) was an important result since this parameter positively affects the price of milk and determines the speed and quality of milk coagulation for cheese making. In pasture based diets, lipid supplementation does not usually affect milk protein concentration <sup>[17,63]</sup> while in confined feeding systems this parameter is systematically affected <sup>[62,64]</sup>. Inclusion of LiOi in the ration of dairy cows does not seem to affect milk protein content or yield <sup>[34,36,44,54]</sup>. In confined conditions, supplementation with unprotected lipids negatively affected milk protein content in 71% of the cases analyzed by <sup>[53]</sup> and the result was also associated with a reduction in casein synthesis <sup>[65,66]</sup>. A large number of studies demonstrated a negative effect of supplemental lipids on the protein concentration of bovine milk [56,60,62,67]. The effect seems more consistent with the use of saturated fats (-0.18 g of protein/100g of milk) and calcium salts of FA (-0.12 g of protein/100g of milk) with respect to unsaturated vegetable oils <sup>[62]</sup>. From the analysis of 8 supplementation trials feeding unsaturated lipids to grazing dairy cows, a decrease (-3.2%) in milk protein concentration (-0.11 g/100g) was reported <sup>[17]</sup>. The physiological mechanisms that explain this reduction are not fully elucidated. Some alteration in ruminal fermentation that reduces microbial protein synthesis and therefore amino-acid availability for the mammary gland for milk protein synthesis has been proposed [70]. A dilution effect after milk production increase has also been suggested <sup>[17,62,64,70]</sup>. In the present work, the increased levels of the oil-blend intake did not affect milk production, milk protein content or yield (Table 4).

Lactose content was also not affected (P > 0.05) after intake of the oil-mixture (Table 4). The apparent decrease in *de novo* mammary lipogenesis (Table 7) implies some reduction in glucose oxidation for NADPH synthesis which could have been spared glucose and increased its bioavailability for lactose synthesis an effect that does not appear to have occurred in the present work. Indeed, plasma circulating glucose levels did not change after oil feeding (Table 5). The lack of changes in milk lactose content would be explained by its osmoregulatory capacity. Some authors suggest that changes in lactose content would only occur in very extreme and infrequent feeding situations <sup>[71]</sup>. Milk urea concentration was not affected by increasing oil intake, a result consistent with the absence of a depressing effect of supplemental lipids on pasture intake (Table 3).

#### **3.3** Concentration of Plasma Metabolites and Changes in Body Weight

Plasma circulating levels of urea, triglycerides, cholesterol, glucose and NEFA were not affected (Table 5).

Table 5. Plasma metabolite concentration in grazing dairycows supplemented or not (Control) with a blend of soy-bean (75%) and linseed (25%) oils at 2%, 4% and 6% ofDM intake.

D		Treat	SEM	<i>P</i> -value <sup>2</sup>		
rarameter	Control	Tr2%	Tr4%	Tr6%		
Urea, mg/dl	45.87	39.50	44.24	48.05	3.88	0.45
Triacylglycerides, Mmol/L	0.25	0.30	0.30	0.31	0.02	0.41
Cholesterol, mg/dl	220.97	235.44	257.1	252.31	14.54	0.29
Glucose, mg/dl	71.92	70.60	69.13	69.38	3.12	0.91
NEFA, µeq/L	276.71	274.93	280.89	335.38	30.94	0.48

*Note:* <sup>1</sup>Values expressed as least squares means and standard error of least squares means (SEM). <sup>2</sup>Treatment effect. NEFA= non-esterified fatty acids.

The absence of negative effects on glycemia suggests that the availability of gluconeogenic precursors was not affected by lipid intake which is compatible with the absence of an isoenergetic replacement of carbohydrates by oil in the concentrate and with the lack of negative effects of the oil-blend on concentrate or total DM intakes (Table 3). When total intake was not affected plasma glucose levels remained constant after protected lipid intake, <sup>[72]</sup>. Even when DM intake was decreased by duodenal infusion of rapeseed oil, glycemia remained unchanged in early or mid-lactation dairy cows<sup>[73]</sup>. The linear reduction observed in concentration of de novo synthesized FA (Table 7) suggests a lower mammary lipogenesis which may have contributed to maintain plasma glucose levels due to a lower oxidation of glucose for NADPH production at the mammary level <sup>[73]</sup>.

Increases in the circulating levels of all plasma lipids after lipid supplementation is a well-documented result <sup>[74,75]</sup> explained by the increase in all fractions of plasma lipoproteins <sup>[65,74]</sup>. The only exception are triacylglycerides due to their high turnover rate <sup>[74]</sup> which could explain the lack of effect of the increasing supply of the oil mixture on triglyceridemia (Table 5).

Plasma NEFA was not affected by supplemental lipids a result that was consistent with the positive LW changes observed in all treatments (Table 6) and also with other experiments <sup>[77,80]</sup>.

**Table 6.** Bodyweight (BW) changes in grazing dairy cows supplemented or not (Control) with a blend of soybean (75%) and linseed (25%) oils at 2%, 4% and 6% of DM intake.

Param-	. Treatments <sup>1</sup>				SEM	<i>P</i> -value			
eter	Control	Tr2%	Tr4%	% Tr6%	SEM	Trat <sup>2</sup>	Lin <sup>3</sup>	Quad <sup>3</sup>	Cub <sup>3</sup>
Initial BW, kg	599.00a	521.50b	556.40ab	538.71b	18.29	0.04	0.50	0.12	0.07
Final BW, kg	647.83a	554.83b	598.80ab	584.00bc	18.78	0.02	0.60	0.05	0.04
Daily BW gain, kg	1.62a	1.12b	1.42b	1.51ab	0.11	0.02	0.40	0.03	0.07
ΔBW, kg	48.83a	33.33b	42.30ab	45.29a	3.41	0.03	0.43	0.01	0.07

*Note:* <sup>1</sup>Values expressed as least squares means and standard error of least squares means (SEM). <sup>2</sup>Treatment effect. <sup>3</sup>Contrasts: linear, quadratic and cubic. <sup>a,b</sup> Means in the same row with different superscripts differ significantly for treatments effect with P-value as mentioned in column for significance at p<0.05 (Test Tukey-Kramer).  $\Delta$ BW=final BW – initial BW.

#### 3.4 Milk Fatty Acid Profile

Compared to Control, milk concentration of butyric acid ( $C_{4:0}$ ) resulted lower (P < 0.05) only after the maximum oil-blend dose at Tr6% (Table 7). Concentration of  $C_{4:0}$  is generally not affected by lipid intake since it is synthesized by an independent malonyl-CoA pathway and therefore not associated with the acetyl-CoA carboxylase activity that is inhibited by the uptake of exogenous FA from oils <sup>[1,81]</sup>. Any reduction in the levels of  $C_{4:0}$  in milk is undesirable for its beneficial effects on human health <sup>[3]</sup>. In this context, intake of the oil-blend at 4% of the total DM would be the maximum recommended dose.

With respect to Control, the decrease in the concentration of *de novo* synthesized MF-A ( $C_{4:0}$  to  $C_{15:1}$ ) was significant only from the Tr4% dose (Table 7). The results reported in <sup>[6]</sup> also showed a reduction (-22.4%) in the total *de novo* synthesized MF-A from 21.07 to 16.35 g/100g when the same oil mixture was fed at 0.8 Kg/cow/day to grazing dairy cows. A reduction in total concentration of *de novo* synthesized FA from 22.49 to 18.48 g/100g of total MF-A (-18.8%) was aldso reported in grazing dairy cows that consumed 0.7 kg/cow/day of a 70:30 blend of

SoOi and LiOi<sup>[80]</sup>. These effects are explained by the inhibition of the activity of the mammary lipogenic enzymes such as acetyl-CoA carboxylase<sup>[65,76]</sup> and are normally reported when dairy cows are supplemented with sources of PUFA<sup>[82,83]</sup>. The inhibitory effect becomes more potent as the length of the PUFA chain and the degree of unsaturation increases and with the presence of double bonds of *trans* configuration<sup>[43]</sup>.

Since at the higher dose (Tr6%) of the oil-blend intake milk fat content was affected (Table 4), the inclusion of the Tr4% dose would be suitable in order to maintain the commercial milk value in a context of payment by quantity of useful (fat-protein) milk solids. At higher oil doses, the decrease in mammary de novo MF-A synthesis did not appear to be compensated by a proportional increase in preformed exogenous FA uptake and milk fat content decreased (Table 4). Milk fat depression was maximal in Tr6% with the highest *trans*-10  $C_{18:1}$  concentration in milk fat (Table 7). This trans-10 isomer has showed deleterious effects on human health [84] and is negatively correlated with milk fat concentration (Figure 1). A high trans-10  $C_{18:1}$  content or the isomer *trans*-10, *cis*-12  $C_{18:2}$  CLA in milk has been related to dysfunctions in the activity of the lipoprotein lipase (LPL) and stearyl CoA desaturase (SCD) enzymes involved in fat synthesis thus causing a decrease in milk fat content<sup>[82]</sup>.

Concentration of total hypercholesterolemic MF-A (C<sub>12:0</sub> to C<sub>16:0</sub>) decreased significantly with the Tr4% oil mixture without an additional reduction at the higher Tr6% dose (Table 7). At the same time, the atherogenicity index (AI) of milk decreased at the Tr4% dose without additional detriments (P > 0.05) between the Tr4% (1.18) and the Tr6% (0.95) doses. Feeding 0.8 Kg/cow/day of the SoOi75:LiOi25 mix showed a 41% decrease in the AI compared to the basal value of 1.93 recorded in the Control treatment in <sup>[6]</sup>. In the present work, the basal AI was 1.87 (Table 7) being thus comparable to that observed in <sup>[6]</sup>.

Feeding 0.8 Kg/cow/day of the 75% (SoOi)-25% (LiOi) mix induced significant reductions in the  $C_{12.0}$  (-30.6%),  $C_{14:0}$  (-28.8%) and  $C_{16:0}$  (-21.9%) in the experiment by <sup>[6]</sup>. In the present trial, concentration of  $C_{12:0}$  and  $C_{16:0}$  showed the same response pattern (Table 7). Compared to Control, concentration of myristic ( $C_{14:0}$ ) acid showed a 21.6% reduction at the T4% dose (Table 7) a result of concern due to the putative atherogenic role the  $C_{14:0}$  MF-A <sup>[2]</sup> when consumed in excess. The observed reductions registered in Tr4% dose for milk content of  $C_{12:0}$  (35.9%),  $C_{14:0}$  (21.7%) and  $C_{16:0}$  (15.5%) were slightly lower than the range estimated in the meta-analysis by <sup>[37]</sup> when supplementing with SoOi and LiOi. They reported values of 42-37% for  $C_{12:0-2}$ , 23-24% for  $C_{14:0}$  and 30-17% for  $C_{16:0}$ .

Table 7. Milk fatty acid (MF-A) composi	ition from grazing dairy co	ows supplemented or not (	Control) with a blend of
soybean (75%) and	d linseed (25%) oils at 2%	6, 4% and 6% of Total DM	I.

MF-A		Treatment <sup>1</sup> <i>P</i> -value				ue			
g/100g MF-A	Control	Tr2%	Tr4%	Tr6%	SEM	Treat <sup>2</sup>	Lin <sup>3</sup>	Quad <sup>3</sup>	Cub <sup>3</sup>
C <sub>4:0</sub>	2.81 <sup>a</sup>	2.36 <sup>ab</sup>	2.33 <sup>ab</sup>	1.84 <sup>b</sup>	0.21	0.02	0.02	0.92	0.35
C <sub>6:0</sub>	1.73 <sup>a</sup>	1.40 <sup>a</sup>	1.21 <sup>ab</sup>	0.91 <sup>b</sup>	0.13	0.001	0.001	0.91	0.65
C <sub>8:0</sub>	1.03 <sup>a</sup>	0.85 <sup>a</sup>	0.64 <sup>b</sup>	0.45 <sup>b</sup>	0.08	<.0001	<.0001	0.96	0.85
C <sub>10:0</sub>	2.49 <sup>a</sup>	2.12 <sup>a</sup>	1.46 <sup>b</sup>	1.08 <sup>b</sup>	0.18	<.0001	<.0001	0.97	0.51
C <sub>12:0</sub>	3.04 <sup>a</sup>	2.77 <sup>a</sup>	1.95 <sup>b</sup>	1.58 <sup>b</sup>	0.19	<.0001	<.0001	0.81	0.28
C <sub>14:0</sub>	10.90 <sup>a</sup>	10.50 <sup>a</sup>	8.54 <sup>b</sup>	7.09 <sup>c</sup>	0.40	<.0001	<.0001	0.20	0.26
C <sub>14:1 cis-9</sub>	1.21 <sup>a</sup>	1.35 <sup>a</sup>	0.92 <sup>ab</sup>	0.95 <sup>ab</sup>	0.12	0.03	0.01	0.96	0.05
C <sub>16:0</sub>	24.99ª	24.55 <sup>a</sup>	21.11 <sup>b</sup>	20.51 <sup>b</sup>	0.72	0.0002	<.0001	0.91	0.08
C <sub>16:1</sub>	0.89	1.23	0.95	1.17	0.13	0.22	0.97	0.67	0.07
C <sub>17:0</sub>	0.45 <sup>a</sup>	0.59 <sup>a</sup>	0.53 <sup>ab</sup>	0.29 <sup>ac</sup>	0.06	0.01	0.09	0.005	0.94
C <sub>18:0</sub>	11.50	9.67	11.58	11.06	0.69	0.20	0.29	0.34	0.06
C <sub>18:1</sub> Isomers									
trans-9	0.21 <sup>a</sup>	0.40 <sup>b</sup>	0.64 <sup>c</sup>	0.73 <sup>d</sup>	0.05	<.0001	<.0001	0.32	0.43
trans-10	0.20 <sup>a</sup>	$0.40^{b}$	0.91°	2.23 <sup>d</sup>	0.17	<.0001	<.0001	0.003	0.52
trans-11 (VA)	3.63 <sup>a</sup>	4.97 <sup>b</sup>	7.05°	8.38 <sup>d</sup>	0.24	<.0001	<.0001	0.98	0.18
Total trans	4.04 <sup>a</sup>	5.77 <sup>b</sup>	8.60°	11.35 <sup>d</sup>	0.34	<.0001	<.0001	0.15	0.45
<i>cis</i> -9 C <sub>18:1</sub>	25.74 <sup>a</sup>	26.04 <sup>a</sup>	27.90 <sup>a</sup>	30.06 <sup>b</sup>	0.20	0.005	0.002	0.28	0.74
<i>cis</i> -11 C <sub>18:1</sub>	1.57 <sup>a</sup>	1.36 <sup>a</sup>	1.94 <sup>ab</sup>	1.91 <sup>ab</sup>	0.170	0.06	0.01	0.60	0.07
C <sub>18:2</sub> (n-6)	1.98 <sup>a</sup>	2.60 <sup>b</sup>	2.81 <sup>b</sup>	2.78 <sup>b</sup>	0.20	0.03	0.02	0.12	0.88
C <sub>18:3</sub> (n-3)	0.58	0.78	0.66	0.64	0.06	0.40	0.73	0.18	0.28
<i>cis-</i> 9, <i>trans-</i> 11 C <sub>18:2</sub> CLA (RA)	2.28ª	3.16 <sup>b</sup>	3.88 <sup>c</sup>	3.89°	0.22	<.0001	<.0001	0.05	0.57
Short chain MF-A <sup>4</sup>	8.06 <sup>a</sup>	6.72 <sup>a</sup>	5.63 <sup>ab</sup>	4.27 <sup>bc</sup>	0.55	0.0005	0.0002	0.98	0.83
Medium chain MF-A <sup>5</sup>	43.33ª	42.67 <sup>a</sup>	34.99 <sup>b</sup>	32.48 <sup>b</sup>	1.09	<.0001	<.0001	0.40	0.02
Long chain MF-A <sup>6</sup>	48.14 <sup>a</sup>	50.03 <sup>a</sup>	58.40 <sup>b</sup>	62.33°	1.32	<.0001	<.0001	0.44	0.08
Saturated MF-A (SMF-A)	60.69 <sup>a</sup>	52.38 <sup>b</sup>	47.59°	43.38 <sup>d</sup>	1.15	<.0001	<.0001	0.08	0.57
Unsaturated MF-A (UMF-A)	38.52ª	42.93 <sup>b</sup>	48.69 <sup>c</sup>	53.39 <sup>d</sup>	1.04	<.0001	<.0001	0.90	0.64
SMF-A/UMF-A	1.59 <sup>a</sup>	1.23 <sup>b</sup>	0.98°	0.82 <sup>cd</sup>	0.06	<.0001	<.0001	0.12	0.96
AI <sup>7</sup>	1.87ª	1.64 <sup>a</sup>	1.18 <sup>b</sup>	0.95 <sup>bc</sup>	0.09	<.0001	<.0001	0.99	0.26
Δ9D products	34.96 <sup>a</sup>	38.22 <sup>b</sup>	44.30°	49.02 <sup>d</sup>	1.06	<.0001	<.0001	0.49	0.40
Substrates <sup>8</sup>	55.65ª	54.41 <sup>a</sup>	52.51 <sup>b</sup>	52.29 <sup>bc</sup>	0.65	0.003	0.0005	0.44	0.43
Índex <sup>9</sup>	0.38ª	0.41 <sup>b</sup>	0.46 <sup>c</sup>	0.48 <sup>d</sup>	0.009	<.0001	<.0001	0.95	0.40
De novo MF-A (C <sub>4:0</sub> to C <sub>15:1</sub> )	24.76 <sup>a</sup>	22.82 <sup>a</sup>	17.90 <sup>b</sup>	14.70 <sup>c</sup>	1.01	<.0001	<.0001	0.53	0.31
Preformed MF-A (>C <sub>17:0</sub> )	48.89 <sup>a</sup>	50.61 <sup>a</sup>	58.93 <sup>b</sup>	62.62°	1.33	<.0001	<.0001	0.46	0.07
Ω6/Ω3	3.57 <sup>a</sup>	3.37 <sup>a</sup>	4.41 <sup>b</sup>	4.63°	0.32	0.02	0.003	0.52	0.16
RA/VA	0.56ª	0.59 <sup>a</sup>	0.49 <sup>b</sup>	0.37°	0.03	<.0001	<.0001	0.01	0.32
$\sum (C_{12:0} \text{ to } C_{16:0})$	38.93 <sup>a</sup>	37.81ª	31.59 <sup>b</sup>	29.18 <sup>b</sup>	1.15	<.0001	<.0001	0.57	0.10

*Notes:* <sup>1</sup> Values expressed as least squares means and standard error of least squares means (SEM). <sup>2</sup>Treatment effect. <sup>3</sup>Contrasts: lineal y cuadratic and cubic. <sup>a,b</sup> Means in the same row with different superscripts differ significantly for treatments effect with P-value as mentioned in column for significance at P<0.05 (Test Tukey-Kramer). <sup>4</sup>Short chain MF-A (C<sub>6.0</sub> to C<sub>10.0</sub>). <sup>5</sup>Mediun chain MF-A (C<sub>12.0</sub> to C<sub>17.1</sub>). <sup>6</sup>Long Chain MF-A (C<sub>18.0</sub> to C<sub>22.6</sub>). <sup>7</sup>Atherogenicity index (C<sub>12.0</sub> + 4\*C<sub>14.0</sub> + C<sub>16.0</sub>/( $\sum$ UMF-A). UMF-A = cis-9 C<sub>14.1</sub>, C<sub>16.1</sub>, cis-9 C<sub>18.1</sub>, cis-11 C<sub>18.1</sub>, trans-11 C<sub>18.1</sub>, C<sub>18.2</sub>, cis-9, trans-11C<sub>18.2</sub> (CLA). The detrimental MF-A *trans*-6, 8, 9, 10 C<sub>18.1</sub> was excluded. <sup>8</sup>Substrates: C<sub>14.0</sub> + C<sub>15.0</sub> + C<sub>16.0</sub> + C<sub>17.0</sub> + C<sub>18.0</sub> + *trans*-11C1<sub>8.1</sub>. <sup>9</sup>Index: ([ $\sum \Delta$ 9D poducts]/[ $\sum \Delta$ 9D products + Substrates]).

The level of stearic acid ( $C_{18:0}$ ) did not differ (P > 0.05) from Control in any of the oil-blend doses used with a significant and linear (P < 0.01) increase of linoleic acid ( $C_{18:2}$ ). These results could be explained by a possible inhibition in the biohydrogenation from  $C_{18:2}$  to  $C_{18:0}$  when high levels of linoleic acid are present in runen <sup>[86,88]</sup>. The absence of increases in milk content of  $C_{18:0}$  after supplementation with oils rich in  $C_{18:2}$  or  $C_{18:3}$  can be considered a positive result due to its potential thrombogenic role <sup>[2]</sup> and was consistent with other experiments [69,80].

The level of oleic acid (*cis*-9 C<sub>18:1</sub>) increased (P < 0.05) over Control only in Tr6% at maximum dose of the oil mixture (Table 7). Using the same SoOi-LiOi mixture at a dose of 0.8 kg/cow/day, the oleic acid content (g/100g MF-A) in Control milk (26.14) did not differ from that observed in supplemented cows (27.50) <sup>[6]</sup>. This result was consistent with that obtained in the Tr4% treatment (Table 7). Differences in oleic acid levels were also not detected

after feeding 0.7 Kg/cow/day of a 70% (SoOi)-30% (LiOi) mixture to grazing dairy cows <sup>[80]</sup>. However, the increase in milk oleic content after supplementation with sunflower or SoOi oils is a frequently reported result <sup>[37,44,89]</sup> even when LiOi is fed <sup>[35-37,90]</sup>.

The linear (P < 0.0001) increase in the desaturation index used to estimate mammary desaturation activity (Table 7) was compatible with the increase in oleic acid, contributing in part to maintaining similar levels of C<sub>18:0</sub> in milk (Table 7). A higher desaturase index was reported in the milk of animals supplemented with LiOi despite the fact that PUFA feeding inhibits the activity of the  $\Delta$ 9-D enzyme complex <sup>[45]</sup>. Several authors did not observe differences in this index when comparing a control ration with those that included SoOi and LiOi or their mixtures at 50-50 <sup>[36]</sup>.

Milk fat content of linoleic acid increased (P < 0.02) linearly from 1.98 (Control) to 2.78 g/100g MF-A in Tr6% (Table 7) thus remaining within the normal range (2-3 g/100g MF-A) reported by <sup>[1]</sup> but lower than that observed in our previous experiment (3.25 to 3.92 g/100g MF-A) after supplementation with 0.7 Kg/cow/day of a mixture of 70% (SoOi)-30% (LiOi) in grazing cows <sup>[80]</sup>. Likewise, an increase in milk C<sub>18:2</sub> content from 1.96 (Control) to 3.50 g/100g MF-A was reported after intake of the same SoOi LiOi mixture at 0.8 Kg/cow/day <sup>[6]</sup>.

On the other hand, the levels of linolenic acid ( $C_{18:3}$ ) in milk did not differ (P > 0.05) between treatments (Table 7), a result consistent with that observed in <sup>[80]</sup>. Feeding LiOi at 4% of DM intake in pure form increased (170%) the levels of  $C_{18:3}$  in milk a result not observed when cows were supplemented with a 50% mixture with SoOi <sup>[36]</sup>.

The milk  $\Omega 6/\Omega 3$  ratio from Control cows was low (3.57) and increased (P < 0.02) with the increasing supply of the oil mixture but always remaining below the recommended value of 5. When the same oil mixture was fed at 4% of total DM intake to grazing dairy cows, the  $\Omega 6/\Omega 3$  ratio remained between 3.18 in control milk and 3.87 in supplemented cows <sup>[80]</sup>. In order to achieve a low  $\Omega 6/\Omega 3$  ratio, the supply of LiOi alone at 4% of total DM intake was very effective averaging 2.13 in supplemented vs. 4.25 in control cows. The effect was attenuated when using mixtures with SoOi <sup>[6]</sup>. In that work <sup>[6]</sup>, the milk  $\Omega 6/\Omega 3$  ratio after intake of 0.8 Kg/cow/day of the 75% (SoOi)-25% (LiOi) blend was greater (5.66) tan observed in the present experiment.

Feeding non-protected PUFA oils increase synthesis of differents isomers of *trans*- $C_{18:1}$  MF-A in the rumen which are transferred to milk. Some of them, like the *trans*-9 (elaidic) and *trans*-10  $C_{18:1}$ , are classed as deleterious or unhealthy <sup>[85,91,92]</sup> and hence any excessive increase in

concentration in milk should be avoided. Basal concentration (g/100g MF-A) of trans-9 (0.21) and trans-10  $C_{18:11}$  (0.20) was linearly (P < 0.01) increased after oilblend intake (Table 7) reaching máximum values of 0.73 for trans-9 and 2.23 for trans-10 C<sub>18-1</sub> in Tr6% that can be considered low or harmless. Indeed, a concentration of 2.28 g/100g MF-A of trans-10 C<sub>18:1</sub> in a butter supplied at 12% of the diet of experimental rabbits subjected to a cholesterol challenge did not show deleterious effects on the plasma lipid profile or the metabolism of lipoproteins when the level of *trans*-10  $C_{18:1}$  was accompanied by at least 7 g/100g MF-A of VA and 3 of RA [85]. In Tr4%, the trans-9 (0.64 g/100g of total MF-A) and trans-10  $C_{18:1}$ (0.91 g/100g MF-A) concentrations in milk fat were lower (P < 0.05) than in Tr6% showing some advantage. Since in Tr4%, concentrations of trans-10 C<sub>181</sub> were only 0.91 g/100g MF-A of the total MF-A with levels of VA and RA in milk of 7.05 and 3.88 g respectively (Table 7) it can be expected an athero-protective role of Tr4% milk similar (or even higher) to that obtained in  $^{[85]}$ . Intake of 0.7 kg/ cow/day of a 70% (SoOi)-30% (LiOi) blend also induced low values of trans-9 (0.58 g/100g of total MF-A) and trans-10 C<sub>18:1</sub> (0.99 g/100g of total MF-A) in milk from grazing dairy cows [80]. In Control milk, concentration of VA accounted for 90% of the total trans-C<sub>18:1</sub> MF-A a value that remained high (74 to 86%) after intake of the oil-blend doses (Table 7). In Control milk, trans-9 and trans-10 C<sub>18:1</sub> represented 5.19 and 4.95% of total *trans*- $C_{18:1}$  but the relative contribution of the *trans*-10 C<sub>18:1</sub> increased after feeding the oil-blend. The relative increase expressed as % of the total *trans*- $C_{18-1}$ , resulted greater in treatments Tr4% (10.5%) and Tr6% (19.7%). Although the concentrations of these two trans FA were moderate, it is convenient to avoid deviations towards its formation due to its potential atherogenic effect [84,85]. It seems therefore advisable not to use doses greater than 4% of supplementary lipids in order to avoid non-undesirable deviations towards non-healthy trans isomers appearance.

The concentration of VA in milk showed a linear increase (P < 0.01) after increasing the oil mixture intake but its apparent conversion into RA estimated by the RA/ VA ratio showed an opposite ratio reaching a minimum value of 0.37 in Tr6% (Table 7). The result suggests that the increase in precursor availability (VA) for RA synthesis did not induce proportional increases in the activity  $\Delta$ -9D desaturase which was also consistent with the lack (P> 0.05) of increase in milk RA content between treatments Tr4% (3.88 g/100g MF-A) and Tr6% (3.89 g/100g MF-A).

An average RA/VA ratio of 0.41 was proposed by <sup>[93]</sup> a value that resulted close to the 0.49 observed in Tr4%

(Table 7). VA present in milk and dairy products can exert beneficial properties in itself through a direct anticarcinogenic effect <sup>[94]</sup> or mediated after its endogenous conversion to RA at an estimated rate of 20% in human tissues <sup>[95]</sup> via  $\Delta$ 9-desaturase activity <sup>[93]</sup>.

Since only 20% of the VA would be converted into RA in human tissues and until more experimental evidence of the healthy effects of VA is available, it seems advisable to avoid excess milk VA concentration and intake. In this context, our results suggest that the oil-blend fed at 4% of total DM intake (Tr4%) would be the most advisable dose since marginal increases in RA were not detected when VA increased with higher oil-blend intake up to Tr6% (Ta-ble 7 and Figure 2).



**Figure 2.** Concentration of vaccenic acid (*trans*-11 C<sub>18:1</sub>) in milk from grazing dairy cows supplemented with increased levels of a soybean-linseed oil blend (75:25)

As expected, milk RA concentration correlated positively ( $R^2 = 0.89$ , P < 0.01) with VA (Figure 3), a frequently reported result <sup>[1,36]</sup>. The average conversion rate of VA into RA appeared to be 34.2% (Figure 3) that resulted very close to the 35-39% values estimated at <sup>[80]</sup> after supplying 700 g/cow/day of a mixture of 70% (SoOi)-30% (LiOi) blend to grazing dairy cows. Taking the RA/VA ratio as an estimator, the average conversión rate resulted somehaw higher (50.25%, Table 7).



**Figure 3.** Concentration of vaccenic acid (*trans*-11 C<sub>18:1</sub>) in milk from grazing dairy cows supplemented with increased levels of a soybean-linseed oil blend (75:25)

In the present work, the baseline (Control) concentration of RA resulted very high (2.28 g/100g MF-A) and was increased (P < 0.05) after oil-blend intake without differences (P > 0.05) in RA concentration between Tr4% and Tr6% (Table 7). This result suggests that the response in milk RA content would be linear up to a maximum of 4% of oil-blend consumption (Figure 4) and confirms what was previously suggested by <sup>[1]</sup>.



**Figure 4.** Concentration of rumenic acid (RA, *cis*-9 *trans*-11  $C_{18:2}$ ) in milk from grazing dairy cows supplemented with increased levels of a blend of soybean and linseed oil (75:25)

At a similar oil-dose (4% DM intake), milk concentration of RA observed in Tr4% (3.88 g/100g MF-A) resulted higher than those reported in <sup>[36]</sup> (1.60-2.39 g/100g MF-A) in rations with a high forage content (59%) and also than those obtained in <sup>[6]</sup> using the 75% (SoOi)-25% (LiOi) oil blend (3.21 g/100g MF-A). When grazing dairy cows were fed 0.7 Kg/cow/day of a mixture of 70% (SoOi)-30% (LiOi), milk concentration of RA averaged 3.13 g/100g MF-A <sup>[80]</sup>.

Milk content of RA in Tr4% (Table 7) was also higher than values reported in the meta analysis by <sup>[37]</sup> after feeding SoOi (1.02 ( $\pm$  0.36) g/100g MF-A) or LiOi (1.75 ( $\pm$ 0.84) g/100g MF-A) and that those reported in <sup>[44]</sup> when the cows were supplemented with 0.5 kg/day of sunflower or soybean oil (2.02 g/100g MF-A) or after feeding 0.9 kg/cow/day of unsaturated FA calcium salts <sup>[98]</sup>. The high baseline values of RA observed in Control milk (2.28 g/100g MF-A, Table 7 and Figure 3) could partly explain the differences between experiments.

The presence of unsaturated MF-A increased linearly (P < 0.0001) with oil intake reaching a maximum at Tr6% with a 38.6% increase over Control (Table 7). The SMF-A/UMF-A ratio decreased (P < 0.0001) with oil intake without differences between Tr4% and Tr6% (Table 7). The results obtained confirmed the existence of great response plasticity in the composition of milk fat in terms of its constitutive MF-A<sup>[1,37]</sup> which can be modulated by oil supplementation to increase the healthy value of dairy products.

#### 4. Conclusions

Supplementation of grazing dairy cows with increasing doses of a mixture of soy and linseed oils linearly increased the rumenic and vaccenic acid content of milk without significant deviations towards unhealthy fatty acids like *trans*-9 or *trans*-10 C<sub>18:1</sub> and without affecting either milk production or protein content. This nutritional strategy was also an effective tool to reduce milk content of saturated milk fatty acids and the hypercholesterolemic fraction of milk fat which improves its healthy value. Overall results shows that the optimum level of inclusion of the soybean-linseed oil mixture was around 3.91% of total DM intake of cows without additional advantages by increasing the dose of oils in the total ration.

#### Acknowledgments

This work was supported by the National Institute of Agricultural Technology (INTA). This Institute is a decentralized state agency with operational and financial autarchy, under the Ministry of Agroindustry of the Argentine Republic. This publication is part of the requirements to access to the academic degree of Doctor in Agricultural Sciences by the Mar del Plata National University, Argentina.

#### References

- Chilliard Y., Ferlay A., Mansbridge, R.M., Doreau, M. Ruminant Milk Fat Plasticity: Nutritional Control of Saturated, Polyunsaturated, trans and Conjugated Fatty Acids. Annales de Zootechnie, 2000, 49: 181-205.
- [2] Ulbright, T.L.V., Southgate, D.A.T. Coronary Heart Disease: Seven Dietary Factors. Lancet, 1991, 338: 985-992.
- [3] Tholstrup, T., Ehnholm, C., Jauhiainen, M., Petersen, M., Hoy, C.E., Lund, P., Sandstrom, B. Effects of Medium-Chain Fatty Acids and Oleic Acid on Blood Lipids. Lipoproteins, Glucose, Insulin and Lipid Transfer Protein Activities. American Journal Clinical Nutrition, 2004, 79, 564-569.
- [4] Parodi, P.W. Conjugated Linoleic Acid and Other Anticarcinogenic Agents of Bovine Milk Fat. Journal of Dairy Science, 1999, 82: 1339-1349.
- [5] Gagliostro, G.A. Nutritional Control of Conjugated Linoleic Acid (CLA) Content in Milk and in Natural and Functional Foods. 2. Production of CLA-Enriched Milk in the Dairy Cow. Revista Argentina de Producción Animal, 2004, 24 (3-4): 137-163.
- [6] Antonacci, L.E., Gagliostro, G.A., Cano, A.V., Bernal, C.A. Effects of Feeding Combinations of Soy-

bean and Linseed Oils on Productive Performance and Milk Fatty Acid Profile in Grazing Dairy Cows. Agricultural Sciences, 2017, 8. 984-1002. https://doi.org/10.4236/as.2017.89072

- [7] Kritchevsky, D., Tepper, S.A., Wright, S., Tso., P., Czarnecki, S.K. Influence of Conjugated Linoleic Acid (CLA) on Establishment and Progression of Atherosclerosis in Rabbits. Journal American College Nutrition, 2000, 19: 472S-477S.
- [8] Valeille, K., Ferezou, J., Amsler, G., Quignard-Boulange, A., Parquet, M., Gripois, D., Dorovska-Taran, V., Martin, J.C. A *cis-9*, *trans-11-Conjugated Linole*ic Acid-Rich Oil Reduces the Outcome of Atherogenic Process in Hyperlipidemic Hamster. American Journal of Physiology Heart and Circulating Physiology, 2005, 289: H652-659.
- [9] Gagliostro, G.A. Nutritional Control of Conjugated Linoleic Acid (CLA) Content in Milk and in Natural and Functional Foods. 1. Effects on Human Health. Revista Argentina de Producción Animal, 2004, 24 (3-4): 113-136.
- [10] Stachowska, E. Conjugated Dienes of Linoleic Acid and Tumorigenesis. Annales of Academic Medical Stetin, 2008, 54: 122-125.
- [11] Shiraishi, R., Iwakiri, R. Fujise, T., Kuroki. T. Kakimoto, T., Takashima, T. Sakata, Y., Tsunada, S., Nakashima, Y., Yanagita, T., Fujimoto, K. Conjugated Linoleic Acid Suppresses Colon Carcinogenesis in Azoxymethane-Pretreated Rats with Long-Term Feeding of Diet Containing Beef Tallow. Journal of Gastroenterology, 2010, 45: 625-635.
- [12] Kim, J. H., Kim, Y. Kim, Y.J., Park, Y. Conjugated Linoleic Acid: Potential Health Benefits as a Functional Food Ingredient. Annual Review of Food Science and Technology, 2016, 7: 221-244.
- [13] Kelly, G.S. Conjugated Linoleic Acid: a Review. Alternative Medicine Review, 2001, 6: 367-382.
- [14] Bhattacharya, A., Rahman, M.M., McCarter, R., O'Shea, M., Fernandes, G. Conjugated Linoleic Acid and Chromium Lower Body Weight and Visceral Fat Mass in High-Fat-Diet-Fed Mice. Lipids, 2001, 41: 437-444.
- [15] Stanton, C., Murphy, J., McGrath, E., Devery, R. Animal Feeding Strategies for Conjugates Linoleic Acid Enrichment of Milk. In: Advances in Conjugated Linoleic Acid in Food. J.L Sébédio. W.W. Christie. R. Adloff (Eds.). AOCS Press. Champaign. Illinois. 2003, 2: 123-145.
- [16] Gagliostro, G.A. Garciarena, D.A., Rodriguez, M.A., Antonacci, L.E. Feeding Polyunsaturated Supplements to Grazing Dairy Cows Improve the Healthy Value of Milk Fatty Acids. Agricultural Sciences,

2017, 8: 759-782.

https://doi.org/10.4236/as.2017.88057

- [17] Schroeder, G.F., Gagliostro, G.A., Bargo, F., Delahoy, J.E., Muller, L.D.. Effects of Fat Supplementation on Milk Production and Composition by Dairy Cows on Pasture: a Review. Livestock Production Science, 2004, 86: 1-18.
- [18] Fernandez, H.H. Estimation of Herbage Allowance. Course Principles of Nutrition and Supplementation in Grazing Cattle. INTA EEA Balcarce, 1999.
- [19] Meijs, J.A.C., Walters, R.J.K., Keen, A. Sward Methods. In: Herbage Intake Handbook. British Grassland Society, 1982: 11-36.
- [20] Horneck, A. D., Miller, R. O. Determination of Total Nitrogen in Plant Tissue. In: Karla. Y. P. Ed: Handbook of Reference Methods of Plants Analysis CRC. Press. 1998: 75-83.
- [21] Morris, L.D. Quantitative Determination of Carbohydrates with Dreywood's Anthrone Reagent. Science, 1948, 107: 254-255.
- [22] Komarek, A.R., Robertson, J.B., Van Soest, P.J. Comparison of the Filter Bag Technique to Conventional Filtration in the van Soest NDF Analysis of 21 Feeds. Proceeding National Conference on Forage Quality. Evaluation and Utilization. Fahey, G.C. Jr. (Ed). Nebraska University. Lincoln. NE. 1994: 02.
- [23] Komarek, A.R. An Improved Filtering Technique for the Analysis of Neutraldetergent Fiber and Acid Detergent Fiber Utilizing the Filter Bag Technique. Journal of Animal Science, 1993, 71: 824-829.
- [24] AOAC. Official Methods of Analysis of the Association of Official Agricultural Chemists. 18th Edn. (1st revision). AOAC International, Gaithersburg. MD. USA, 2006.
- [25] McRae, J.E., Armstrong, D.G. Enzyme Method for Determination of Alpha-Linked Glucose Polymers in Biological Materials. Journal of Science and Food Agriculture, 1968, 19: 578-581.
- [26] AOAC. Official Methods of Analysis of the Association of Official Agricultural Chemists. 18th Edn. (1st revision). AOAC International. Gaithersburg. MD. USA,1999.
- [27] Wolff, R. L. Content and Distribution of trans 18:1 Acids in Ruminant Milk and Meat Fats. Their Importance in European Diets and their Effect on Human Milk. Journal of American Oil and Chemistry Society, 1995, 72: 259-272.
- [28] Bannon, C. D., Breen, G. J., Craske, J. D., Hai. N. T., Harper, N. L., O'Rourke, K. L. Analysis of Fatty Acid Methyl Esters with High Accuracy and Reliability. III. Literature Review of and Investigations into the Development of Rapid Procedures for the Me-

thoxide Catalysed Methanol of Fats and Oils. Journal of Chromatography, Amsterdam, 1982, 247(1): 71-89.

[29] Masson, L. T., Alfaro, C., Camilo, A., Carvalho, P., Illesca, R., Torres, M., Tavares do Carmo, J., Mancini-Filho, A., Bernal, C.A. Fatty Acid Composition of Soybean/Sunflower Mix oil, Fish Oil and Butterfat Applying the AOCS Ce 1j-07 Method with a Modified Temperature Program. Grasas y Aceites, 2015, 66: e064. ISSN-L: 0017-3495. DOI: http://dx.doi.org/10.3989/

ISSN-L: 0017-3495. DOI: http://dx.doi.org/10.3989/ gya.0692141

- [30] SAS/STAT® User's Guide. 2002-2010 by SAS Institute Inc. Cary. NC. USA.
- [31] Minson, D.J. Forage in Ruminant Nutrition. Academic Press Inc., San Diego, California. 1990: 482.
- [32] Leaver, J.D. Herbage Intake Handbook. British Grassland Society. Hurley 1982: 143.
- [33] Verité, R. and Journet, M. Influence of Water Content and Dehydration of the Forage on Its Dietary Value for Dairy Cows. Annales de Zootechnie, 1970, 10: 269-277.
- [34] Loor, J.J., Ferlay, A., Ollier, A., Doreau, M., Chilliard, Y. Relationship Among trans and Conjugated Fatty Acids and Bovine Milk Fat Yield due to Dietary Concentrate and Lindseed Oil. Journal of Dairy Science, 2005, 88: 726-740.
- [35] Flowers, G., Ibrahim, S.A., AbuGhazaleh, A.A. Milk Fatty Acid Composition of Grazing Dairy Cows when Supplemented with Linseed oil. Journal of Dairy Science, 2008, 91: 722–730.
- [36] Bu, D.P., Wang, J.G., Dhiman, T.R., Liu, S.J. Effectiveness of Oils Rich in Linoleic and Linolenic Acids to Enhance Conjugated Linoleic Acid in Milk from Dairy Cows. Journal of Dairy Science, 2007, 90: 998-1007.
- [37] Glasser, F., Ferlay, A., Chilliard, Y. Oilseed Lipid Supplements and Fatty Acid Composition of Cow Milk: a Meta-Analysis. Journal of Dairy Science, 2008, 91: 4687-4703.
- [38] Martinez, M.G. Modulation of the Fatty Acid Composition of Bovine and Caprine Milk through Supplementation with Soybean and Fish Oil. Master Science Thesis, Faculty of Agrarian Sciences, National University of Mar de Plata, Argentina, 2010: 130.
- [39] Martínez del Olmo, D. Supplementation of Rations for High Production Dairy Cows with Oils of Plant Origin: Productive and Reproductive Yields. PhD thesis. Complutense University of Madrid. Veterinary school. Animal Production Department. 2012: 122. (in Spanish)
- [40] Dewhurst, R.J., Shingfield. K.J., Lee. M.R.F., Scol-

lan, N.D. Increasing the Concentrations of Beneficial polyunsaturated Fatty Acids in Milk Produced by Dairy Cows in High-Forage Systems. Animal Feed Science and Technology, 2006, 131: 168-206. DOI: 10.1016/j.anifeedsci.2006.04.016

- [41] Jenkins, T.C. Feeding Fat to Dairy Cattle. In: Proceedings of the Dairy Herd Management Conference. University of Georgia. Athens. CA., 1994: 100-109
- [42] Sutton, J.D., Knight, R., McAllan, A.B., Smith, R.H. Digestion and Synthesis in the Rumen of Sheep Given Diets Supplemented with Free and Protected Oils. British Journal of Nutrition, 1983, 49: 419-432.
- [43] Chilliard, Y., Ferlay, A., Doreau, M. Effect of Different Types of Forages. Animal Fat or Marine Oils in Cow's Diet on Milk Fat Secretion and Composition, Especially Conjugated Linoleic Acid (CLA) and Polyunsaturated Fatty Acids. Livestock Production Science, 2001, 70: 31-48.
- [44] Rego, O.A., Rosa, H.J.D., Portugal, P., Cordeiro. R., Borba, A.E.S., Vouzela, C.M., Bessa, R.J.B. The Effects of Supplementation with Sunflower and Soybeans Oils on the Fatty Acid Profile of Milk Fat from Grazing Dairy Cows. Animal Research, 2005, 54: 17-24.
- [45] Chilliard, Y., Martin, C., Rouel. J., Doreau, M. Milk Fatty acids in Dairy Cows fed Whole Crude Linseed, Extruded Linseed, or Linseed Oil, and their Relationship with Methane Output. Journal of Dairy Science, 2009, 92: 5199–5211.
- [46] Pantoja, J., Firkins, J.L., Eastridge, M.L. Fatty Acid Digestibility and Lactation performance by Dairy Cows Fed Fats Varying in Degree of Saturation. Journal of Dairy Science, 1996, 79: 429-437.
- [47] Kim, E.J., Huws, S.A., Lee, M.R.F., Wood, J.D., Muetzel, S.M., Wallace, R.J., Scollan. N.D. Fish Oil Increases the Duodenal Flow of Long Chain Polyunsaturated Fatty Acids and *trans*-11 18:1 and Decreases 18:0 in Steers via Changes in the Rumen Bacterial Community. Journal of Nutrition, 2008, 138: 889-896.
- [48] Atkinson, R.L., Toone, C.D., Robinson, T.J., Harmon, D.L., Ludden, P.A. Effects of Ruminal Protein Degradability and Frequency of Supplementation Nitrogen Retention, Apparent Digestibility and Nutrient Flux Across Visceral Tissues in Lambs Fed Low-Quality Forage. Journal of Animal Science, 2009, 10: 2246-2257.
- [49] Dschaak, C. M., Noviandi, C. T., Eun, J.-S., Fellner, V., Young, A. J., ZoBell, D. R. and Israelsen, C. E. Ruminal Fermentation, Milk Fatty Acid Profiles, and Productive Performance of Holstein Dairy Cows fed 2 Different Safflower Seeds. Journal of Dairy Sci-

ence, 2011, 94: 5138-5150.

- [50] Doreau, M., Chilliard, Y. Effects of Ruminal or Postruminal Fish Oil Supplementation on Intake and Digestion in Dairy Cows. Reproduction Nutrition Dévelopmènt, 1997, 37: 113–124.
- [51] Ueda, K., Ferlay, A., Chabrot, J., Loor, J. J., Chilliard, Y., Doreau. M. Effect of Linseed Oil Supplementation on Ruminal Digestion in Dairy Cows Fed Diets with Different Forage: Concentrate Ratios. Journal of Dairy Science, 2003, 86: 3999–4007.
- [52] Palmquist, D. L. Use of Fats in Diets for Lactating Dairy Cow. Fats in Animal Nutrition. Butterworths. London. 1984: 357-381.
- [53] Morand-Fehr, P., Chilliard, Y., Bas, P. Impact of Including Fats in the Ration on Yield and Composition of Ruminant Milk. Institut National de la Recherche Agronomique, 1986, 64: 59-72.
- [54] Rego, O. A., Alves, S. P., Antunes. L.M. S., Rosa, H. J. D., Alfaia C. F. M., Prates, J. A. M., Cabrita, A. R. J., Fonseca, A. J. M., Bessa, R. J. B. Rumen Biohydrogenation-Derived Fatty Acids in Milk Fat from Grazing Dairy Cows Supplemented with Rapeseed, Sunflower, or Linseed Oils. Journal of Dairy Science, 2009, 92: 4530–4540.
- [55] Dhiman, T.R., Satter, L.D., Pariza, M.W., Galli, M.P., Albright, K., Tolosa, M.X.. Conjugated Linoleic Acid (CLA) Content of Milk from Cows Offered Diets Rich in Linoleic and Linolenic Acid. Journal of Dairy Science, 2000, 83: 1016–1027.
- [56]Alzahal, O., Odongo, N.E., Mutsvanqwa, T., Or-Rashid, M.M., Duffield, T.F., Baqq, R., Dick. P, Vessie. G., McBride, B.W. Effects of Monensin and Dietary Soybean Oil on Milk Fat Percentage and Milk Fatty Acid Profile in Lactating Dairy Cows. Journal of Dairy Science, 2008, 91 (3): 1166-1174.
- [57] Huang, Y., Schoonmaker, J.P., Bradford, B.J., Beitz, D.C. Response of Milk Fatty Acid Composition to Dietary Supplementation of Soy Oil, Conjugated Linoleic Acid, or Both. Journal of Dairy Science, 2008, 91: 260–270.
- [58] Piperova, L.L., Teter, B.B., Bruckental, I., Sampugna, J., Mills, S.E., Yurawecz., M.P., Fritsche, J. Ju, K, Erdman, R.A. Mammary Lipogenic Enzyme Activity, trans Fatty Acids and Conjugated FattyAacids are Altered in Lactating Dairy Cows fed a Milk-Fat Depressing Diet. Journal of Nutrition, 2000, 130: 2568-2574.
- [59] Whitlock, L.A., Schingoethe, D.J., AbuGhazaleh, A.A., Hippen, A.R., Kalscheur, K.R. Milk Production and Composition from Cows Fed Small Amounts of Fish Oil and Extruded Soybean. Journal of Dairy Science, 2006, 89: 3972-3980.

- [60] Shingfield, K. J., Reynolds, C. K., Hervas, G., Griinari, J. M., Grandison, A. S., Beever, D.E. Examination of the Persistency of Milk Fatty Acid Composition Responses to Fish Oil and Sunflower Oil in the Diet of Dairy Cows. Journal of Dairy Science, 2006, 89: 714–732.
- [61] Chilliard. Y., Ferlay, A., Loor, J., Rouel. J., Martin, B. Trans and Conjugated Fatty Acids in Milk from Cows and Goats Consuming Pasture or Receiving Vegetable Oils or Seeds. Italian Journal of Animal Science, 2002, 1: 243-254.
- [62] Gagliostro, G.A., Chillard, Y. Protected Lipid Utilization in Dairy Cows Nutrition. I. Effects on Milk Production and Composition, Dry Matter and Energy Intake (A Review). Revista Argentina de Producción Animal, 1992, 12, (1): 1-15.
- [63] Bargo, F., Muller, L.D., Kolver, E.S., Delahoy, J.E. Invited Review: Production and Digestion of Supplemented Dairy Cows on Pasture. Journal of Dairy Science, 2003, 86: 1-42.
- [64] Wu, Z., Huber, J.T. Relationship Between Dietary Fat Supplementation and Milk Protein Concentration in Lactating Cows: A Review. Livestock Production Science, 1994, 39: 141-155.
- [65] Storry, J.E. The Effect of Dietary Fat on Milk Composition. In: Recent Advances in Animal Nutrition. W. Haresing. (Eds). Butterworths. London, 1981: 3-33.
- [66] Dunkley, W.L., Smith, N.E., Franke, A.A. Effect of Feeding Protected Tallow on Composition of Milk and Milk fat. Journal of Dairy Science, 1977, 60: 1863-1869.
- [67] Palmquist, D.L., Beaulieu, A.D., Barbano, D.M. Feed and Animal Factors Influencing Milk Fat Composition. Journal of Dairy Science, 1993, 76: 1753–1771.
- [69] Ward, A. T., Wittenberg, K. M., Przybylski, R. Bovine Milk Fatty Acid Profiles Produced by Feeding Diets Containing Solin, Flax and Canola. Journal of Dairy Science, 2002, 85: 1191–1196.
- [70] Garnsworthy, P.C. Fats in Dairy Cow Diets. In: Recent Advances in Animal Nutrition. pp 87–13. Cole. D.J.A. (eds.). Nottingham UniversityPress. Nottingham. UK, 1997.
- [71] Vicente, G.R., Shelford, J.A., Peterson, R.G., Khrishnamurti, C.R. Effects of Feeding Canola-Meal-Protected Tallow or Soy-Bean-Meal-Protected Tallow in the Low-Roughage Diet of Dairy Cows in Early Lactation. Canadian Journal of Animal Science, 1984, 64: 81.
- [72] Gagliostro, G.A., Chilliard, Y., Davicco, M.J. Duodenal Rapeseed Oil Infusion in Early and Mid-Lactation Cows. 3. Plasma Hormones and Mammary Apparent Uptake of Metabolites. Journal of Dairy

Science, 1991, 74: 1893-1903.

- [73] Chilliard, Y., Gagliostro, G.A., Fléchet. J., Lefaivre, J., Sebastian, I. Duodenal Rapeseed Oil Infusion in Early and Mid Lactation Cows. 5. Milk Fatty Acids and Adipose Tissue Lipogenic Activities. Journal of Dairy Science, 1991, 74: 1844-1854.
- [74] Christie, W.W. The Effects of Diet and Other Factors on the Lipid Composition of Ruminant Tissues and Milk. In: Lipid metabolism of ruminant animals. Christie W.W. (Ed). Pergamon Press. Oxford, 1981: 193-226.
- [75] Mandebvu, P., Ballard, C.S., Sniffen, C.J., Carter, M.P., Wolford, H.M., Sato, T., Yabuuchi, Y., Bolck, E., Palmquist, D.L. Effect of Feeding Calcium Salts of Long-Chain Fatty Acids from Palm Fatty Acid distillate or Soybean Oil to High Producing Dairy Cows on Milk Yield and Composition and on Selected Blood and Reproductive Parameters. Animal Feed Science and Technology, 2003, 108: 25-41.
- [76] Ambrose. D. J., Kastelic, J. P., Corbett, R., Pitney, P.A., Petit, H.V., Small, J.A., Zlkovic, P. Lower Pregnancy Losses in Lactating Dairy Cows Fed a Diet Enriched in Linoleic Acid. Journal of Dairy Science, 2006, 89: 3066–3074.
- [77] Castañeda-Gutierrez, E., Benefield, B.C., de Veth, M.J., Santos, N.R.. Gilbert, R.O. Butler, W.R., Bauman, D.E. Evaluation of the Mechanism of Action of Conjugated Linoleic Acid Isomers on Reproduction in Dairy Cows. Journal of Dairy Science, 2007, 90: 4253-4264.
- [78] Fuentes, M.C., Calsamiglia, S.S., Sánchez, C., González, A., Newbold, J., Santos, J. E.P., Rodríguez-Alcalá, L.M., Fontecha, J. Effect of Extruded Linseed on Productive and Reproductive Performance of Lactating Dairy Cows. Livestock Production Science, 2008, 113: 144-154.
- [79] Chilliard, Y., Ferlay, A. Dietary Lipids and Forages Interactions on Cow and Goat Milk Fatty Acid Composition and Sensory Properties. Reproduction Nutrition Dévelopmènt, 2004, 44: 467–492.
- [80] Antonacci, L., Rodríguez, A., Castelli, L., Zampatti, M., Castañeda, R., Ceaglio, J., Gagliostro, G.A. Suplementación con Mezcla de Aceites Vegetales y el Perfil en Acidos Grasos de Leche Bovina. Revista Argentina de Producción Animal. 2013, 33(1).
- [81] Casper, D.P., Shingoethe, D.J., Middaugh, R.P., Baer, R.J. Lactational Responses of Dairy Cows to Diets Containing Regular and High Oleic Sunflower Seeds. Journal of Dairy Science, 1988, 71: 1267–1274.
- [82] Bauman, D.E., Griinari, J.M. Regulation and Nutritional Manipulation of Milk Fat: Low-Fat Milk Syndrome. Livestock Production Science, 2001, 70:

15-29.

- [83] Kelly, M.L., Berry, J.R., Dwyer, D.A., Griinari, J.M., Chouinard, P.Y., Van Amburgh, M.E., Bauman, D.E. Dietary Fatty Acid Sources Affect Conjugated Linoleic Acid (CLA) Concentrations in Milk from Lactating Dairy Cows. Journal of Nutrition, 1998, 128: 881–885.
- [84] Roy, A., Ferlay, A., Chilliard, Y. Production of Butter Fat Rich in *trans*-10C18:1 for Use in Biomedical Studies in Rodents. Reproduction Nutrition Dévelopmènt, 2007, 46: 211-218.
- [85] Hartoof, C.G., Noble, R.C., Moore, J. H. Factors Influencing the Extent of Biohydrogenation of Linoleic Acid by Rumen Micro-Organisms *in vitro*. Journal of Science and Food Agriculture, 1973, 24: 961–970.
- [86] Agazzi, A., Bayourthe, C., Nicot, M.C., Troegeler-Meynadier, A., Moncoulon, R., Enjhalnbert, M. In Situ Ruminal Biohydrogenation of Fatty Acids from Extruded Soybeans: Effects of Dietary Adaptation and of Mixing with Lecithin or Wheat Straw. Animal Feed Science and Technology, 2004, 117: 165-175.
- [87] Cruz-Hernandez, C., Kramer, J.K.G., Kennelly, J.J., Glimm, D.R., Sorensen, B.M., Okine, E.K., Goonewardene, L. A., Weselake, R. J. Evaluating the Conjugated Linoleic Acid and Trans 18:1 Isomers in Milk Fat of Dairy Cows Fed Increasing Amounts of Sunflower Oil and a Constant Level of Fish Oil. Journal of Dairy Science, 2007, 90: 3786–3801.
- [88] Hurtaud, C., Faucon, F., Couvreur, S, Peyraud, J.L. Linear Relationship Between Increasing Amounts of Extruded Linseed in Dairy Cow Diet and Milk Fatty Acid Composition and Butter Properties. Journal of Dairy Science, 2010, 93: 1429–1443.

- [89] Mozaffarian, D., Katan, M. B., Ascherio, A., Stampfer, M. J., Willett, W.C., Food, T. Trans Fatty Acids and Cardiovascular Disease. New England Journal of Medicine, 2006: 1601-1613.
- [90] Dorfman, S.E., Laurent, D., Gounarides, J.S., Li, X., Mullarkey, T.L., Rocheford, E.C., Sari-Sarraf, F. Metabolic Implications of Dietary *trans*-fatty acids. Obesity, 2009, 17: 1200-1207.
- [91] Chilliard, Y., Gasser, G., Enjalber, F., Ferlay, A., Bocquier, F., Schimidely, P.H. Conference: Recent Results on the Effects of Feeding on the Fatty Acid Composition of Cow's Milk. Goat and Sheep. Argentine Journal of Animal Production, 2007, 27(3): 197-213. (in Spanish)
- [92] Awad, A.B., Hermann, T., Fink, C.S., Horvath, P.J. 18:1 n7 Fatty Acids Inhibit Growth and Decreased Inositol Phosphate Release in HT-29 Cells Compared to n-9 Fatty Acids. Cancer Letters, 1995, 91: 55-61.
- [93] Turpeinen, A.M., Mutanen, M., Aro, A., Salminen, I., Basu, S., Palmquist, D.L., Griinari, J.M. Bioconversion of Vaccenic Acid to Conjugated Linoleic Acid in Humans. American Journal of Clinical Nutrition, 2002, 76: 504-510.
- [94] Parodi, P.W., Conjugated Linoleic Acid in Food. In: Advances in Conjugated Linoleic Acid in Food. J.L Sébédio. W.W. Christie. R. Adloff (Eds.). AOCS Press. Champaign. Illinois, 2003, 2: 101-122.
- [95] Griinari, J.M., Bauman, D.E. Biosynthesis of Conjugated Linoleic Acid and its Incorporation Into Meat and Milk in Ruminants. In: M. P. Yurawecz. M. M. Mossoba. J. K. G. Kramer. M. W. Pariza and G. J. Nelson (Eds.) Advances in Conjugated Linoleic Acid Research. AOCS Press. Champaign, 1999: 180-200.



**Journal of Zoological Research** http://ojs.bilpublishing.com/index.php/jzr



#### ARTICLE

# Five years Retrospective Study of Avian Coccidiosis in a Veterinary Clinic Bukuru Plateau State Nigeria

Barde, Israel Joshua<sup>1\*</sup> Ladan, Haruna Bello<sup>2</sup> Shekaro, Audu<sup>1</sup> Ijoma, Sandra Ifynneke<sup>1</sup> Idachaba, Stella Ejura<sup>1</sup> Olabode Victoria Bose<sup>1</sup> Oguche, Moses Ojonugwa<sup>1</sup> Ishaku, Bata Shalangwa<sup>2</sup>

1. National Veterinary Research Institute, Vom, Plateau State, Nigeria

2. Federal College of Animal Health and Production Technology, Vom, Plateau State, Nigeria

#### ARTICLE INFO

Article history Received: 15 June 2020 Accepted: 27 June 2020 Published Online: 30 June 2020

*Keywords:* Avian coccidiosis Plateau state Retrospective study Veterinary clinic

#### ABSTRACT

This study was conducted to evaluate the incidence of avian coccidiosis and its associated various risk factors such as age, type of birds and season in a private veterinary clinic in Bukuru, Plateau State Ngeria. A total of 9406 cases during 2013 - 2017 were analysed and 1556 of them were positive for coccidiosis. There are several reports on the prevalence of avian coccidiosis by previous researchers; however, in this study we evaluated the prevalence of avian coccidiosis in the study area and its economic impacts. Total prevalence of 12.14% in 2013, 18.78% in 2014, 18.21% in 2015, 16.82% in 2016 and 19.07% in 2017 were reported. An overall prevalence of 85.02% was recorded. The average prevalence of coccidiosis based on this five years study is 17%. The association between coccidiosis and age of the birds was determined and age 5-8 weeks becomes most effective period with wet season having high percentage prevalence of coccidiosis. Based on the type of birds, coccidiosis is prevalence almost in equal proportion in both broilers and lavers. The losses caused by avian coccidiosis could be both direct and indirect components which may include the cost of control measures, inadequate good hygiene practices, production losses and lack of prophylaxis treatment. The control of avian coccidiosis can be achieved through good sanitary measures by avoiding water spillage on the pen floor, overcrowded stocking density, the use of prophylaxis- anticoccidials and proper good vaccination practices.

#### **1. Introduction**

The business of poultry farming in Nigeria has in recent times witnessed enormous expansion <sup>[1]</sup>. The estimated commercial poultry birds population in Nigeria rose from 110 million at the beginning of

this century, to over 150 million in 2006 <sup>[6]</sup>. In spite of this enormous growth, the poultry industry has suffered a lot of constraints of which poultry disease is a major player in this setback/contraint <sup>[5]</sup>. Avian coccidiosis in poultry is caused by several species of the genus Eimeria <sup>[16]</sup>. The disease causes reduced growth, emaciation, anaemia

\*Corresponding Author: Barde, Israel Joshua, National Veterinary Research Institute, Vom, Plateau State, Nigeria; Email: israelbarde@yahoo.com and mortality in the infected birds <sup>[7]</sup>. This could results in heavy economic losses due to the cost of treatment and prevention measures <sup>[17]</sup>. Warm and humid weather provides favourable conditions for the growth and development of infective ooysts <sup>[5]</sup> which could result in a high prevalence of coccidiosis in any tropical location like Bukuru.

Avian coccidiosis is caused by protozoan parasite of genus eimeria species of the family Eimeriadae and order Eucoccidiorida<sup>[3]</sup>. The parasites develop within the intestine of most infected poultry birds. Seven species of eimeria (E. acervulina, E. brunetti, E. maxima, E. mitis, E. necatrix, E. praecox and E. tenella) are recognised that infect chickens <sup>[13,18]</sup>. Despite the fact that coccidiosis is a disease known for plenty years, it is still considered as the most economical important parasitic disease condition affecting poultry production world-wide uptil now <sup>[15,18]</sup>. Coccidiosis has played a major role in economic loses to poultry famers the world wide, for example in 1995 the United Kingdom was estimated to have lost thirty eight million pounds due to avian coccidiosis <sup>[10]</sup>. Many poultry famers have not taken sanitary measures serious in their poultry business so disease like coccidiosis have had its way in destroying such farms badly <sup>[10,15]</sup>. Diagnosis of coccidiosis could be based on the history of bloody diarrhea/feces in infected birds or carcasses/moribund birds, with post mortem and microscopy examination considered as one of the best option to be used to confirm its diagnosis. Post mortem examination of infected poultry carcasses may revealed lesions on the serosal and mucosal surfaces of the intestine most times. Some of the lesions observed may include; enteritis of the interior one third, the middle and the posterior one third of the intestine depending on the type of coccidian and these could manifest as hyperemia, necrosis of the intestinal mucosa and bloody feces in the lumen, thus serving as pointer to the presence to the disease, and this could be complimented by microscopy to observe for, gamonts, schizonts and oocysts usually with a lot of successes recorded <sup>[11,17]</sup>. Although prevalence of coccidiois has been reported in various locations in Nigeria by some researchers <sup>[2,9]</sup>, however, in Bukuru Plateau State there is a dearth in data of Eimeria species and the prevalence of coccidiosis. Hence this study seeks to investigate the prevalence of coccidiosis and Eimeria species in Bukuru Plateau state. It is therefore important to survey for the disease that occur most frequently among poultry farmers in Bukuru and environs through this retrospective study. The aim is to conduct a five years retrospective study of avian coccidiosis in a private veterinary clinic Bukuru, Jos south Plateau state during 2013-2017.

#### 2. Materials and Methods

#### 2.1 Study Area

The study was conducted at ECWA veterinary clinic in Bukuru- Nigeria. It is a private clinic which operates in Bukuru near Jos where diagnosis, treatment and other veterinary service are offered for all classes of livestock and pets. The clinic also undertakes vaccination; meanwhile they provide upon request the feasibility studies for different classes of livestock investment. The clinic has a workforce comprising of two veterinary doctors and many attendants, sales men as well as a security man. The clinic under review has a laboratory, post-mortem room, treatment room, surgery room, three quarantine pens, drug and feed stores and three offices.

Bukuru geographical co-ordinates are 90 48' 0' NORTH, 80 52' 0' EAST.

#### 2.2 Methodology

The record of diseases as presented and diagnosed at the ECWA veterinary clinic Bukuru during the five years (2013-2017) period was retrieved and analysed retrospectively. The diagnosis at the clinic was based on clinical signs and post-mortem findings using the method described by Olabode et al.<sup>[13]</sup>.

#### 2.3 Statistical Analysis

Descriptive statistic frequency and percentage was used for categorical variables. Association between infections and other factors such as types of birds, age, etc. were accessed using chi-square test for the association and odd ratio. P<0.05 was considered significant. Descriptive statistic was used to analyse data, tables were used for results and proportions presented in percentages.

#### 3. Results

 
 Table 1. The prevalence of coccidiosis in chickens based on ages

Month of occurrence	1-4 weeks	5-8 weeks	3-6 weeks	> 6months
January	46	32	32	26
February	55	128	121	55
March	43	37	27	27
April	40	46	21	15
May	37	43	22	16
June	57	48	31	19
July	29	36	33	12
August	41	27	22	27
September	40	32	31	25
October	40	49	25	19
November	35	38	27	21
December	35	26	24	25

*Note:* Ages 5-8 weeks and 1-4 weeks appeared to be more infective age. Although at age 3-8 months, the prevalence was also recorded higher but only in Februaries in the 5 years study. While ages above 6-months had least infective periods <sup>[10]</sup>.



Figure 1. Reports of coccidiosis during 2013-2017 based on the age of birds

Season		Occurrence of coccidiosis		Total	
		present	absent		
dry wet or dry	Count	795	3660	4455	
	Expected Count	725.1	3729.9	4455.0	
	Adjusted Residual	3.9	-3.9		
season	eason	Count	731	4190	4921
wet	Expected Count	800.9	4120.1	4921.0	
	Adjusted Residual	-3.9	3.9		
Tota	1	Count	1526	7850	9376
Expected Count		1526.0	7850.0	9376.0	

Table 2. The association between disease and season

*Note:* P<0.05 [P=0.01] df=1 T=15.345 since the P is less than 0.05, it means there is an evidence of significant of association between disease and season in chicken occurring during 2013 to 2017

wet or dry season \* coccidiosis or no coccidiosis Cross tabulation

**Table 3.** Prevalence of coccidiosis during 2013-2017 based on types of birds, (Layers and broilers). Here the infection of coccidiosis is slightly high in the layers than in the broilers with the period under review

BASED ON	TYPES OF	BIRDS FRO	M 2013-2017
MONTHS	LAYERS	BROILERS	
January	64	69	
February	177	182	
March	70	64	
April	64	69	
May	66	53	
June	75	80	
July	58	50	
August	60	51	
Septembe	66	63	
October	60	73	
Novembe	61	44	
Decembe	36	47	
	857	845	



Figure 2. Prevalence of coccidiosis during 2013-2017 based on types of birds

Table 4. The total prevalence of chicken coccidiosis yearly



Figure 3. The total prevalence of coccidiosis yearly in chickens

#### 4. Discussion

Based on results of this study, it was observed that age of the birds plays a vital role in infectivity of coccidiosis which is much prevalent in age 5-8 weeks birds as indicated in table 1 and figure 1. The prevalence showed a decline in birds with age 1-4 weeks, though prevalent in age 3-6months birds was highest but only in the months of February throughout the five years study. This means that age 3-6 months and above were the least infective period which could be due to previous exposure of the old birds. This agrees with the report of Chauham and Sushovan <sup>[4]</sup> and Mark et al. <sup>[10]</sup>. There is an evidence of significant association between the presence of coccidiosis in chicken and the age group occurring within 2013-2017 where the observed value in the adjusted residual value of coccidiosis presence in chicken with age group 1-8 weeks was less than what would have been expected through chance alone. While in the other hand the adjusted residual value the observed value of coccidiosis in chicken with age 1-8 group was more than what would expect. Meanwhile, in the adjusted residual value of coccidiosis presence in chicken with age group of > 24 weeks was found to be less than what would expect. The overall results show that the disease is much prevalent in the younger birds. Chickens within ages 1-8 weeks were most diagnosed with coccidiosis. The highest infective period was between 9-24 weeks of age with great declined at age > 24 weeks old, this is in agreement with Chauhan and Sushovan <sup>[4]</sup> and Mark et al. <sup>[10]</sup>.

Table 2 indicates the association between disease and season (P<0.05). There is a significant association between disease and season in chicken occurring during 2013 to 2017. From the adjusted residual value we had more coccidiosis than expected through chance alone in the dry season' also we had in the wet season less observation than we expected through chance alone. Eimeria oocysts remain viable in litter for many months, hence they contaminate the farm year in year out thereby maintaining infection of the poultry flock. The Eimeria oocysts can killed by freezing and extreme dryness and high temperature, thereby making it difficult for the infection to get spread in the dry and cold month as agreed by <sup>[4,5]</sup>. Also wet area constitutes a source of avian coccidiosis infection. The poultry pen litters are supposed to be applied in thick layers in order to facilitate maximum absorption of the bird's waste droppings. The wet litters enhances sticking of materials to boots, utensils, shoes, vehicle wheels and clothing leading to a faster rate of transfer of the organism to other farms. Bukuru is a wet and damp place hence the prevalence of coccidiosis among poultry farmers as agreed with the point raised by Chauham and Sushovan<sup>[3,4,12]</sup>

Table 3 shows prevalence of coccidiosis during 2013-2017 based on types of birds, (Layers and broilers). Here the infection of coccidiosis is slightly high in the layers than in the broilers within the period under review but figure 2 shows that there is no significance difference in coccidiosis prevalence based on the types of birds. This means that coccidiosis is prevalence almost in equal proportion in both boilers and layers in this study. The observed value presence in broilers was more than the expected and this occur by chance alone while compared to the layers where in the adjusted residual value, the value observed of coccidiosis presence in layers occur was more than the expected and this occur by chance alone. There was no significant difference (p>0.05) between the prevalence rate of coccidiosis in layers and broilers. This is in agreement with the report of Olanrenwaju and Agbor<sup>[14]</sup>. In layer and broiler, the prevalence rate of coccidiosis in birds reared under deep litter system and battery cage system of management shows a strong association between system of management and occurrence of the disease with the former system being higher. The higher prevalence rate in broilers and layers reared under deep litter system of management compared with the battery cage system of management is in agreement with the report of Etuk et al.<sup>[5]</sup>, Hadipour et al.<sup>[7]</sup> and Jatau et al.<sup>[8]</sup>

Table 4 and figure 3 show the total prevalence of coccidiosis was lower in 2013, having the least of 12.14% prevalence, while in 2014 the prevalence elevated to about 18.78%, and then declined a little bit to 18.21% in 2016. Reasons could be because of adequate awareness in hygiene practice in poultry production. This agrees with the point earlier raised by Chauham and Sushovan <sup>[4]</sup>. Subsequently the infection became high on 2017 with increase rate of 19.07% which might be due to decrease in the awareness of good bio-security practice.

#### 5. Conclusion

The average prevalence of coccidiosis in this study was 17 %. Age plays a great role in infectivity of coccidiosis with age 5-8 weeks birds showing much prevalent, with wet season having high prevalence. Coccidiosis had prevalence almost in equal proportion in both boilers and layers in present study. Poultry farmers should adhere to good vaccination and hygienic practices as well as the use of both drugs and vaccines to prevent the incidence of the disease. Farmers should adhere to routine chemoprophylaxis and avoid factors of predisposition to cooccidiosis especially during the raining season. A further study of the genetic basis of parasite survival on the hosts and the key molecules associated with the disease would be essential so as combat the disease effectively.

#### Acknowledgements

Our special acknowledgements go to the Management of ECWA Veterinary clinic Bukuru Nigeria for allowing us access to their clinic records

#### **Competing Interest**

The authors declare that they have no competing interests.

#### Authors' contributions

BIJ and LHB designed the research; ISI, ISE,SA and OVB performed the research and analysed the data; OMO wrote the manuscript; IBS have taken part in the revision of the manuscript. All authors read and approved the final version of the manuscript.

#### References

- Adewole, S.O.The efficacy of drugs in the treatment of coccidiosis in chicken in selected poultries. Academic Research International, 2012, 2(1): 20-24.
- [2] Barde J I, Garba A, Gashua MM, Talba MA, Gugong VT, Sa;adatu I, Owada AH, Konzing L, Awulu SJ, Mohammed MN. Common diseases of poultry in Kaduna State; perspective of a private clinic. Nigerian veterinary journal, 2012, 33 (3): 581-585.
- [3] Chauhan HVS, Sushovan R. Poultry Diseases, Diagnosis and Treatment, 3rd Edition. 2000: 145-167.
- [4] Chauhan HVS, sushovan R. Poultry diseases, diagnosis and treatment 2nd edition, new international[P]. LTD Pub, 2003: 61-79.
- [5] Etuk, E.B., Okoli. I.C., Uko, M.U. Prevalence and management issues associated with poultry coccidiosis in Abak agricultural zone of Akwa Ibom State, Nigeria. International Journal of Poultry Science, 2014, 3(2): 135-139.
- [6] FAO. Food and agriculture organisation of united nation, Rome Quaterly bulletin of statistics, 2000, 1: 12-21.
- [7] Hadipour, M.M., Olyaie, A., Naderi, M., Azad, F. and Nekouie, O. Prevalence of coccidiosis in scavenging native chickens of Shiraz, Iran. African Journal of Microbiology Research, 2011, 5(20): 3296-3299.
- [8] Jatau, I.D., Sulaiman, N.H., Musa, I.W., Lawal, A.L., Okubanjo, O.O., Isah, I., Magaji, Y. Prevalence of coccidia infection and preponderance Eimeria species in free range indigenous and intensively managed exotic chickens during hot-wet season, in Zaria, Nigeria. Asian Journal of Poultry Science, 2012, 6(3): 79-88.
- [9] Majaro, O.M. The epidemiology and economic importance of poultry coccidiosis in Oyo state, Nigeria. Rev. Elev. Med. Vet. Pays Trop., 1980, 33: 377- 379.
- [10] Mark P, Paul F, Mcmulin J, Bradbury M, Dennis JA.

Poultry disease 6th edition. Sounders Elsevier Ltd, 2008: 444-460.

- [11] Moses Gyang Davou, Kumbish, P. R., Barde I. J., Ahmad J.S. Olabode H. O. K, Wungak Y. S. A Retrospective Study on Chicken Coccidiosis in Ilorin, Kwara State, Nigeria. Direct Research Journal of Agriculture and Food Science (DRJAFS), 2015, 3(5): 93-97.
- [12] Muazu, A., Masdoog, A.A., Ngbede, J., Salihu, A.E.,Haruna, G., Habu, A. K., Sati, M.N, Jamilu, H.. Prevalence and identification of species of Eimeria causing coccidiosis within Vom, Plateau State, Nigeria. International Journal of Poultry Science, 2018, 7(9): 917-918.
- [13] Olabode Victoria Bose, Dashe Yakubu Gunya, Umaru Mada Alsea, Tobias Peter Pwajok Choji and Israel Joshua Barde. Histopathological Lesions of Coccidiosis Natural Infestation in Chickens. Asian Journal of Research in Animal and Veterinary Sciences, 2020, 5(2): 41-45.
- [14] Olanrenwaju, C.A., Agbor, R.Y.Prevalence of coccidiosis among poultry birds slaughtered at Gwagwalada main market, Abuja, FCT, Nigeria. The International Journal of Engineering and Science, 2014, 3(1): 41-45.
- [15] Safiullin RT, Kachanova EO, Chalysheva EI, Andreyanov ON. Experimental Model of Coccidiosis Caused by Eimeria Tenella in Broiler Chickens. World Veterinary Journal, 2019, 9(4): 262-267.
- [16] Taylor, M.A., Coop, R.L., Wall, R.L.. Veterinary Parasitology (3rd ed). Blackwell Publishing. 2007: 224-234.
- [17] Usman. J.G., Gadzama. U.N., Kwagha. A.V., Madziga. H.A.: Anti-coccidial resistance in poultry: A review. New York Science Journal, 2011, 10(10): 102-106
- [18] William RB. A compartmentalised model for the estimation of the cost of coccidiosis to the world's chicken production industry. International journal for parasitology, 1999, 29 (8): 209-229.



**Journal of Zoological Research** http://ojs.bilpublishing.com/index.php/jzr



# ARTICLE Contribution of Livestock Production to Global Greenhouse Gas Emission and Mitigation Strategies

#### Ahmedin Abdurehman Musa<sup>\*</sup>

Department of Animal Science, Oda Bultum University, P.O. Box 226, Chiro, Oromiya, Ethiopia

ARTICLE INFO	ABSTRACT
Article history Received: 15 June 2020 Accepted: 27 June 2020 Published Online: 30 June 2020	Understanding the interaction of livestock production and climate change is currently the main issue in global warming. This paper reviews the contribution of livestock production in greenhouse gas emission and its mitigation strategies. The potential contribution of individual large rumi- nants are 200-500 litters of methane per day while small ruminants pro- duces 20-40 litters of methane per day. The major greenhouse gas related
Keywords: Livestock Methane Emission Mitigation	to livestock production are methane and nitrous oxide which contribute approximately about 14.5% global GHG emissions. Limiting emissions from livestock, without compromising food security, is an important limit greenhouse gas emissions. The main choices for reducing greenhouse gas emission in livestock production are more related to improving animal production. Mitigating emission of CH4 by means of improved manage- ment of biogas and manure, reducing CH4 emission from enteric fermen- tation through improved efficiency and diet, husbandry as well as genetic management are some of strategies used in mitigating enteric emission of methane from livestock. The other one is mitigating emission of nitrous oxide through more efficient use of nitrous fertilizer, proper manure man- agement and by using different feed additives.

#### 1. Introduction

gricultural production potential has previously grown-up 2.1-2.3 in over the last 40 years as reported by <sup>[1]</sup> and this is responsible for 10-12% of the global anthropogenic greenhouse gas emission <sup>[2]</sup>.

Fossil fuels are the major causes of climate change as first list. Some or the main source of resulted by human related activities as emission of carbon dioxide and some of greenhouse gas are: natural gas, Oil and especially coal. However, the life of animal and animal production as food for human are comprehended that as a main source of greenhouse gases, and this in fact not less than half of human caused greenhouse gases emission [3].

According to the report of <sup>[4]</sup> there is an expectation of Human population increment from 7.2 - 9.6 billion by the year 2050. This indicates that 33% population increase, but obviously as global living standard increase, the increment in demand for agricultural product will increase by around 70% in the future in the same period estimated by <sup>[1]</sup>. Livestock is among one of the fastest growing from agricultural subsectors in developing countries. In these country GDP is around 33% share of the total agricultural GDP and is rapidly increasing. This easily shows that progress is induced by the quickly increasing demand for animal products driven by population growth as well as in-

Ahmedin Abdurehman Musa,

<sup>\*</sup>Corresponding Author:

Department of Animal Science, Oda Bultum University, P.O. Box 226, Chiro, Oromiya, Ethiopia; Email: ahmedin133@gmail.com

creasing income and urbanization in developing countries <sup>[5]</sup>. Ruminants are expected to be an important component of global production and there is a growing demand especially for animal protein sources <sup>[6]</sup>, still keeping their indispensable role in the management and preservation of ecosystems, namely in natural and semi-natural grasslands and rangelands and agrosilvopastoral systems, among others. But, a substantial upsurge in agricultural production will be required to meet these increasing demand for animal originated protein foods <sup>[2]</sup>. This is an event that is likely to lead to strengthened production practice and following increases in Greenhouse gas emission.

Livestock system have both negative and positive effect on social equity and economic growth, natural resource and public health <sup>[7]</sup>. Livestock produces greenhouse gases in different forms like: in the form of nitrous oxide (N<sub>2</sub>O) from the use of nitrogen containing fertilizer, methane (CH<sub>4</sub>) from enteric fermentation and N<sub>2</sub>O and CH<sub>4</sub> from livestock manure deposition on the pastureland and from different animal manure management. Carbon dioxide is also produced on different livestock farms from different energy usage and fuels <sup>[8]</sup>.

Currently there is a huge rising interest in understanding the linkage between agricultural production especially livestock production and climate change and it has been motivating a significant amount of research <sup>[9]</sup>. Therefore, this paper reviews the livestock sector's contribution to the global climate change and its mitigation strategies.

#### **Objectives**

To describe the contribution of livestock sector on climate change

To clarify and summarize mitigation strategies

#### 2. Contribution of Livestock Production Practises to Climate Change

Livestock system plays significant role in climate change <sup>[10]</sup>. Livestock production directly and indirectly contributes about soil carbon loss in grazing land, deforestation for grazing land and intensive animal feed production, the amount of energy used in cultivating and harvesting feed and processing, transporting dairy products, meat and meat products, live animals and animal feed, gases from animal manure (especially CH<sub>4</sub>) and enteric fermentation and nitrous oxide (N<sub>2</sub>O) releases from the use of nitrogen containing synthetic fertilizers<sup>[11]</sup>. Greenhouse gases most often associated with animal production are methane, nitrous oxide and carbon dioxide <sup>[12, 13]</sup>. Similarly, greenhouse gas emission from agricultural sector that are related to animal production are CH<sub>4</sub> which directly emitted from livestock stomach and manure, while nitrous oxide  $(N_2O)$  emitted from fertilizer applied soil and manure as well as grazed lands as reported by<sup>[14]</sup>.

Different authors approximate the contribution of livestock production on global greenhouse emission with different figures.<sup>[15]</sup> estimate the livetock contribution to global anthropogenic greenhouse gas emission at between 7 - 18% . methane is the most important gas prodused in agriculture <sup>[13]</sup>. Ruminant livestock approximatly can produce 250-500 litter of CH<sub>4</sub> per day. This level of production results in estiation of the contribution large ruminant to global warming that may occur in the next 50-100 years to be less than 2% about 65 % of the livestock production emissions. With respect to activities, feed processing and production and enteric fermentation from ruminants are the two major sources of emissions, contributing 45 % and 39 % of total emissions respectively. Manure storage and processing forms 10 % and the rest is attributed to transportation and animal processing. On product-basis, milk from cows and beef are responsible for the most emissions, contributing 20 % and 41 % of the sector's total greenhouse gas (GHG<sup>[16]</sup>. Majority of the livestock industry emission are in the form of methane (44%), while 29% and 27% are nitrous oxide and carbon dioxide respectively (figure 1) below [2,17,18].



Figure 1. Livestock contribution to global GHG emission

The green plant used up by the livestock instigates from the conversion of atmospheric carbon dioxide to biomass or organic compound. Hence, under the Kyoto Protocol (2005) it is assumed that the amount of consumed carbon dioxide in negative form are equivalent to those emitted by the animals. Therefore, livestock respiration is not counted as a net source of Carbone dioxide emission since they are part of global biological cycle. On the other hand, the animal is thought to be a carbon sink since a fraction of the Carbone consumed is absorbed in the live tissue of the livestock and livestock products like milk and meat<sup>[19]</sup>.

Many authors reported that emission form animal production contribute more greenhouse gas emission to the atmosphere than the entire global transportation sector. Thus, the domestic animal donates indirectly and directly to greenhouse gas emission <sup>[20, 21]</sup>.

#### 2.1 Direct Contribution of Livestock to Greenhouse Gas Emission

Some of the direct emission from animal source include animal physiology, respiration, enteric fermentation and excretion <sup>[22]</sup>.

Emission of CH4 is thought as one of the most significant global issue<sup>[2]</sup>. During feed fermentation and digestion in animals, methane gas is produced as by-product of digestion of structural carbohydrate majorly cellulose due to the action of microorganism (fungus, protozoa and bacteria) in the rumen (figure 2). At the time this digestion of monosaccharide are fermented to CO<sub>2</sub>, H<sub>2</sub> and VFA such as propionate, butyrate and acetate <sup>[23]</sup>. This process releases H<sub>2</sub> while producing VFA and some of the microbial cells comprising energy and essential protein to be made available for the growth of animals in all ruminants, the H<sub>2</sub> is removed through the action a group of microorganism known by methanogenic archaea or methanogens that can gain their energy via combining CO<sub>2</sub> and H<sub>2</sub> to form methane <sup>[24]</sup>. Of course,  $CH_4$  is produced by archaeal microorganism known as methanogen which utilizes predominantly carbon dioxide and hydrogen in the rumen to form CH<sub>4</sub> in the animals, thereby maintaining the lower partial pressure of H<sub>2</sub> in the rumen. Actually this CH<sub>4</sub> production from the production from the rumen archaea result in 2-12% loss of metabolizable energy in the rumen <sup>[16,24]</sup>. According to the report of <sup>[2]</sup> GHG account shows that CH<sub>4</sub> emission from livestock is almost equivalent to the GHG emission from the transportation sector in the case of Australia.



Figure 2. Feed and  $H_2$  reduction in the rumen adopted from <sup>[25]</sup>

# **2.2 Indirect Contribution to Greenhouse Gas** Emission

Indirect emission refers to emission resulted from ma-

nure application, manure storage, farm operation, land application chemical fertilizers and manure treatments, feed crop, transportation, animal product processing and land allocation for livestock production (like: desertification, deforestation and carbon release from cultivated land)<sup>[20]</sup>. Generally, in the case of livestock production indirect emission play a great role in the release of C<sub>2</sub>O to the atmosphere than direct emission<sup>[26]</sup>.

Greenhouse gas emission from animal production in particular and agriculture in general are expected to grow as food production expands to keep pace with a growing world which is expected to reach 8.3 billion by 2030 and 9.1 bill by 2050 as estimated by <sup>[4]</sup>.

In developing country especially in Africa, there is an increase in  $CH_4$  emission resulted from increased livestock production. According to the report of <sup>[27]</sup> there was an estimation that African cattle, sheep and goat produce about 7.8mil tons of  $CH_4$  in 2000 which are likely to increase to 11.1mil tons by 2030. As <sup>[28]</sup> reported that, in case this linear relationship between methane emission and livestock population continue, it could be concluded that global methane emission form animals production may increase 60% by the year 2030. Though, the moderate solution for reducing methane emission from livestock production practices could be changing feeding practices and manure management <sup>[29]</sup>.

#### 3. Livestock Sector GHG Mitigation Strategies

Reducing greenhouse gas emissions from livestock, without conceding food security is therefore clearly an important portion of any international effort to limit greenhouse gas emission overall and their effect on climate system<sup>[30]</sup>.

The main alternatives for limiting GHG emission per unit of animal production: firstly, mitigating emission of  $CH_4$  via improved management of biogas and manure; secondly, reducing  $CH_4$  emission from enteric fermentation especially in ruminant animals (mostly cattle, goat and sheep) via improved feed efficiency; thirdly, mitigating emission of NO<sub>2</sub> through more effectual use of inorganic or nitrogenous fertilizers; fourthly, confiscating carbon and mitigating  $CO_2$  emission by reduction and reversal of deforestation due to agricultural intensification and by restoration of organic carbon to cultivated soil and degraded pasture land or rangeland and fifthly, changing the herd structure through increasing the proportion of monogstric animals like pig and chickens as well as vegetarian fish in the flow of animals grown for human consumption <sup>[31]</sup>.

#### 3.1 Methane Mitigation Strategies

Numerous studies have formulated reduction schemes to

mitigate methane emission. Generally, mitigation can be grouped in to two: basically those targeting manure management and those targeted at enteric fermentation<sup>[23]</sup>.

#### **3.1.1 Methane Mitigation Strategies Aimed at En**teric Fermentation

Diminishing enteric  $CH_4$  emission from ruminant livestock without changing livestock production is needed both as a strategy to reduce global greenhouse gas emission and as means of improving feed conversion efficiency of the individual animals<sup>[32]</sup>.

Some of mitigation strategies can be used to reduce greenhouse gas emission such as the use of some specified chemicals and vaccines, genetic selection and the capture of methane have been proposed, yet dietary management is considered the most promising strategy for the diminution of methane from ruminant animal production system [<sup>33</sup>].

#### (1) Genetic Management

Naturally the potential of animals to produce enteric methane is vary. As the first strategy to reduce methane emission per individual animals is the use of selection or selective breeding with an animal's permitting low methane emission per unit of feed consumed 10% with no negative impact on productivity record <sup>[10]</sup>. Therefore, selecting animals that shows excellent production performance on low quality feds is also another way of reducing CH<sub>4</sub> emission per individual animal product.

Another option to reduce methane emission is the potential of changing rumen microorganism. Currently changing the rumen microbial composition in lambs and calves after weaning towards lowering methane emission in the future adult life is being explored and practically available <sup>[10]</sup>.

#### (2) Dietary Manipulation

Dietary manipulation is also the second strategy to reduce methane emission per individual animals. Harvesting pasture and forage at early maturity stage improves its nutritional content of some soluble carbohydrate and decrease the level of lignin in the plant cell wall thus increases its digestibility <sup>[34]</sup> and also reducing enteric methane emission per unit of digestible dry Matter.

Mechanical processing of feeds like processing via its influence on energy losses, passage rate and digestibility can be an effective enteric methane emission mitigation alternative although it may not be economically feasible in some animal production systems. Providing higher quality forage is also another way of reducing enteric methane emission because it improve digestibility of the feed <sup>[15]</sup>.

Another strategy of dietary manipulation is concentrate supplementation. Addition of small amount of concentrate

to all roughage (natural pasture or forage) is expected to increase animal productivity and reduce greenhouse emission per individual animals<sup>[15]</sup>.

Lipid supplementation is the most reliable and technically acceptable nutritional manipulation used to reduce enteric methane emissions. Nevertheless, its diminution potential is ultimately limited by a restriction on dietary inclusion in order to maintain production efficiency [33]. Similarly, <sup>[15]</sup> reported that dietary lipid are effective in reducing enteric methane emission, but the application of this practice will depends on its cost and its effect on feed intake, production and product composition like milk composition. Reductions of 10-25% may be achieved via the supplementation of dietary lipid or oil to the ration of ruminants <sup>[35]</sup>. Some of the possible mechanism by which added oil can reduce CH<sub>4</sub> emission include: (1) by increasing the amount of energy used to digest fiber (mostly in long chain fatty acids); (2) dry matter intake lowering (if total dietary lipid exceeds 6-7%); (3) via suppuration of methanogens mainly in medium-chain fatty acids; (4) through overpowering of rumen protozoa; and (5) to a ) to a restricted extent via bio-hydrogenation <sup>[6, 35]</sup>. According to the evidence of some researchers, a 1% increase of dietary fat can reduce enteric methane emission between 4 -5% [32, 35]

Grinding grain feed or physical processing of grain feed aimed to improve its digestibility is expected to decrease enteric methane emission intensity <sup>[15]</sup>. Improving quality of diet also result in better animal production performance as well as decreasing methane production in the rumen as measured by decrease in methane emission per unit of animal product <sup>[6]</sup>.

Strategic supplementation of the diet like chemical treatment of low quality feeds or pasture, ration balancing and crop selection for straw quality are effective mitigation strategies, but these technology has been poorly practiced in animal feeding<sup>[15]</sup>.

Dietary Protein management is also a good strategy to reduce methane emission. An increase of protein content of diet or ration can also improve digestibility and reduce overall methane emission per unit of animal product <sup>[36]</sup>.

(3) Husbandry Management

Methane emission from a given farm depends on the number of animals and the emission per head <sup>[24]</sup>. Increasing an individual animal productivity can be a very effective strategy for decreasing GHG emission pee unit of animal product. Reduction of herd size is a good strategy, this would also increase feed availability and productivity of individual animals and the total herd, thus sinking methane emission intensity <sup>[15]</sup>.

Minimizing disease and environmental stressor via an

effective disease causing agent management strategy will improve productivity of the herd and results in reduction of  $CH_4$  emission per unit of animal product as well as in overall herd of the farm <sup>[24]</sup>.

Regarding the age of calves to reach slaughter weight and the number of days the cattle remained on feed in the feedlot to finish weight has effect on the rate of methane emission per animals. To resolve these problem improving animal nutrition and genetics can have a significant impact on GHG emission in beef and other meat animal production system<sup>[15]</sup>. In case of dairy farming, extending lactation period is the main strategy to reduce methane emission because it reduces herd energy demand and replacement rate<sup>[12]</sup>.

(4) Chemical Additives

Some chemicals are used in animal feed for the sake of improving feed digestibility. Recently it is known that some chemical agents such as ionophores (monensin), unsaturated fatty acid, sulphate, nitrate, fumarate and halogenated methane analogues (Bromochloromethane (BCM)) are able to reduce methane production from ruminant animals <sup>[16,25,37]</sup>.

Adding nitrate to the ration result in reduced amount of  $CH_4$  emission because it is converted to ammonium  $(NH_4^+)$  which leaves less  $H_2$  available for methane production. This method may have applicability in place such as Australia and Brazil where nitrate could replace the urea which is added to low quality ration to nutritive value <sup>[10]</sup>.

Bromochromomethane (BCM) is one of the most effective inhibitors and apparently reduce CH4 production by interfering with the Cobamide dependent methyl transferase step of methanogenesis <sup>[38]</sup>. Bromochromomethane (BCM) complexed in cyclodextrin CD; BCM-CD) results in the stained inhibition of CH4 production when fed to ruminants <sup>[39]</sup>. Moreover, an in vitro continuous fermentation system simulating rumen fermentation demonstrated that BCM significantly reduced methane production by (85-90%) and eliminated most methanogens, whereas there was no effect on total production, true digestibility of feed and of feed efficiency of microbial protein synthesis <sup>[40]</sup>.

#### (5) Probiotic Supplements

There are some microbial feed additives that have been developed to improve productivity by directly influencing rumen fermentation <sup>[41]</sup>. Probiotics or direct fed microbial are used in the diet of ruminants to improve the health status, rumen fermentation and ultimately the animal per formance that could also reduce methane emission <sup>[42]. [43]</sup> Reported the use of probiotics in mitigation of methane from ruminants. Probiotics improved productivity by 7 to 8 percent resulting in reduced CH<sub>4</sub> per unit of product in

cattle.

# 3.1.2 Methane Mitigation Targeting Manure Management

The most mitigation alternatives for greenhouse gas emission from stored manure, such as reducing the time of aeration, manure storage and stacking are generally aimed at reducing the time of allowed for microbial fermentation process to occur before land application. This kind of mitigation practices are more effective, but their economic feasibility is uncertain<sup>[15].</sup>

Slurry manur storage	Solid Manure Storage	
Storage temperature	Prevent CH <sub>4</sub> formation	
Manure acidification	Prevent anaerobic conditions	
Reduced storage time	Reduced storage time	
Prevent and repair leakage	Composting	
Improve anaerobic digestion	Reduce manure moisture	
Collect and combust methane	Storage temperature	

Manure acidification

Table 1. Methane mitigation strategy from manure.

*Source:* <sup>[15]</sup>

Cover manure storage

#### 3.2 Mitigating Emissions of Nitrous Oxide

Some of the strategies used for increasing the efficiency of N-Cycle in livestock production system and soil aeration should also lead to reduced  $N_2O$  emission <sup>[4]</sup>.

Diminishing total ration protein contain and supplementing the ration with synthetic amino acid is an effective means of ammonia and N2O mitigation strategies for non-ruminants. Ammonia emission from liquid animal waste or slurry receiving the tannin supplemented diet was 8-49% lower than the control slurry. Tannin also lower ammonia emission by 20% when directly applied to the barn flor and 27% bafter a tannin excreta was applied to the soil <sup>[44]</sup>. In contrary to the economic value of the manure, tannin use can reduce N-release rate from manure and thus affect manure -N availability for plant growth <sup>[15]</sup>.

Salt similarly has some mitigation effect of methane in animal production. Adding salt increase water intake in ruminants, this may force the animals both decreasing urinary nitrogen concetration and encouraging more frequent urination events thus spreading urine more evenly across grazing pasture<sup>[6]</sup>.

Another mitigation strategy is by use of chemicals that inhibit the oxidation of ammonium to nitrate in soil and thereby reducing N<sub>2</sub>O emission from urine <sup>[15]</sup>. Some of Nitrification inhibitors like (Dicyandiamide or 3,4-dimethylpyrazole phosphate ) applied with slurry under simulated Portuguese condition were very efficient in reducing nitrous oxide emission<sup>[45]</sup>.

#### 4. Conclusion

The livestock sector contribute indirectly and directly to greenhouse gas emission. Indirect emission include emission resulting from feed crops, farm operation, manure application, transportation, animal product processing and land use allocation for animal production while direct emission from livestock sources refers to enteric fermentation, excretions and respiration. Greenhouse gases most often associated with animal production are methane, nitrous oxide and carbon dioxide. Around 44% of animal emission are in the form of CH4 while N<sub>2</sub>O represent 29% and CO<sub>2</sub> represent 27%. Livestock contribute to global GHG emission approximately 14.5%. Limiting emissions from livestock, without cooperating food security is an important effort to GHG emission. The main option for reducing GHG emission per unit of livestock production include: mitigating emissions of CH<sub>4</sub> through reducing methane emission from enteric fermentation through improved feed efficiency of individual animal, husbandry as well as genetic management and improved management of biogas and manure. The other one is mitigation emission of N<sub>2</sub>O via more efficient use of nitrogenous fertilizer, proper manure feed management and by using different feed additives.

#### References

- [1] FAO 2009. The state of food and agriculture. Livestock in the Balance (Rome: Food and Agriculture Organization of the United Nations), 2009.
- [2] IPCC 2007. Agriculture. In B. Metz, O. R. P. Davidson, R. Bosch, R. Dave, & L. A. Meyer (eds.) Climate Change 2007: Mitigation (Cambridge, New York: Cambridge University Press), Contribution of Working Group III to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change, 2007: 497+540.
- [3] Robert Goodland, Jeff Anhang, 200669. Livestock and Climate Change. UNEP (United Nations Environment Programme). Global environment outlook, 2012 5: Chapter 5. http://www.unep.org/geo/pdfs/geo5/GEO5\_report\_ C5. pdf
- [4] UN, 2008. World Population Prospects: The 2008 Revision. New York, NY, USA.

www.esa.un.org/unpp

[5] Delgado, C. Rising demand for meat and milk in developing countries: implications for grasslands-based livestock production. In Grassland: a global resource (ed. D. A. McGilloway) The Netherlands: Wageningen Academic Publishers, 2005: 29-39.

- [6] Henry, Beverley, Richard Eckard. Greenhouse Gas Emissions in Livestock Production Systems. Tropical Grasslands, 2009, 43:232-38.
- [7] World Bank. Minding the stock: bringing public policy to bear on livestock sector development. Report no. 44010-GLB.Washington, DC, 2009.
- [8] F.P. O'Mara. The significance of livestock as a contributor to global greenhouse gas emissions today and in the near future. Animal Feed Science and Technology, 2011, 166-167: 7-15.
- [9] Aydinalp, C., Cresser, M.S.. The effects of climate change on agriculture. Agric. Environ. Sci. 5, 672-676.
- [10] SAI. 2014. Reducing Greenhouse Gas Emissions from Livestock : Best Practice and Emerging Options. SAI Platform, 2008.
- [11] US EPA. Global anthropogenic non-CO2 greenhouse gas emissions: 1990-2020. Washington DC: US Environmental Protection Agency, Office of Atmospheric Programs, 2006.
- [12] Smith, p., martino, d., cai, z., gwary, d., janzen, h., kumar, p., mccarl, b., ogle, s., o'mara, f., rice, c., scholes, b., sirotenko, o. Agriculture. In: b. Metz, o.r. davidson, p.r. bosch, r. Dave and l.a. meyer (eds) climate change 2007: mitigation. Contribution of working group iii to the fourth assessment report of the intergovernmental panel on climate change. (cambridge university press: cambridge, united kingdom and new york, usa), 2007.
- [13] Sarkwa, F. O., E. C. Timpong-Jones, N. Assuming-Bediako, S. Aikins, and T. Adogla-Bessa. The Contribution of Livestock Production to Climate Change: A Review. Livestock Research for Rural Development, 2016.
- [14] Kebreab, E, K Clark, C Wagner-Riddle, J France. Methane and Nitrous Oxide Emissions from Canadian Animal Agriculture: A Review. 2006.
- [15] Hristov, Alexander N., Joonpyo Oh, Chanhee Lee, Robert Meinen, Felipe Montes, Troy Ott, Jeff Firkins. Mitigation of Greenhouse Gas Emissions in Livestock Production- A Review of Technical Options for Non-CO2 Emissions. FAO Animal Production and Health, 2013, Paper No. 177.

https://doi.org/10.1016/j.agee.2005.08.009

[16] Johnson, K A, D E Johnson. Methane Emissions from Cattle. Journal of Animal Science, 1995, 73: 2483-92.

https://doi.org/10.2527/1995.7382483x

[17] Gerber, P.J., Steinfeld, H., Henderson, B., Mottet, A., Opio, C., Dijkman, J., Falcucci, A., Tempio, G.. Tackling Climate Change Through Livestock: A Global Assessment of Emissions and Mitigation Opportunities. FAO, Rome, 2013.

- [18] Rojas-Downing, M. Melissa, A. Pouyan Nejadhashemi, Timothy Harrigan, Sean A. Woznicki. Climate Change and Livestock: Impacts, Adaptation, and Mitigation. Climate Risk Management 16. The Authors, 2017: 145-63.
- https://doi.org/10.1016/j.crm.2017.02.001 [19] United Nations Framework Convention on Climate
- [19] Onned Pations Planework Convention on Climate Change (UNFCCC). Kyoto Protocol reference manual on accounting of emissions and assigned amount. Climate Change Secretariat (UNFCCC), Martin-Luther-King-Strasse 8, 53175 Bonn, Germany, 1998.
- [20] Havlik, P., H. Valin, M. Herrero, M. Obersteiner, E. Schmid, M. C. Rufino, A. Mosnier, et al. Climate Change Mitigation through Livestock System Transitions. Proceedings of the National Academy of Sciences, 2014, 111 (10): 3709-14. https://doi.org/10.1073/pnas. 1308044111
- [21] Casey, Kenneth D, José R Bicudo, David R Schmidt, Anshu Singh, Susan W Gay. Air Quality and Emissions from Livestock and Poultry Production/Waste Management Systems, 2006.
- [22] Jungbluth, T., Hartung, E., Brose, G., Greenhouse gas emissions from animal houses and manure stores. Nutr. Cycl. Agroecosyst, 2001, 60: 133-145.
- [23] Hopkins, A., A. Del Prado. Implications of Climate Change for Grassland: Impacts, Adaptations and Mitigation Options. Grass and Forage Science, 2007, 62: 118-26.
- https://doi.org/10.1111/j.1365-2494.2007.00575.x
  [24] Henry, B, R Eckard.. Greenhouse Gas Emissions in Livestock Production Systems. Tropical Grasslands, 2009, 43: 232-38.
- [25] Morgavi, D P, E Forano, C Martin, C J Newbold. Microbial Ecosystem and Methanogenesis in Ruminants. The Animal Consortium, 2010, 4 (7): 1024-36. https://doi.org/10.1017/S1751731110000546
- [26] Steinfeld, H., Wassenaar, T., Jutzi, S.. Livestock production systems in developing countries: status, drivers, trends. Rev. Sci. Tech. Off. Int. Epiz., 2006, 25(2): 505-516.
- [27] Herrero, M., Thornton, P.K., Kruska, R., Reid, R.S.. Systems dynamics and the spatial distribution of methane emissions from African domestic ruminants to 2030. Agric. Ecosyst. Environ., 2008, 126: 122-137.
- [28] Bruinsma, J.. World Agriculture: Towards 2015/2030: An FAO Perspective. Earthscan, London, 2003.
- [29] Thornton, P.K., Herrero, M., 2010. The Inter-linkages between rapid growth in livestock production,

climate change, and the impacts on water resources,

- [30] NZAGRC. The Impact of Livestock Agriculture on of Climate Change The Impact Livestock Agriculture on Climate Change, 2012: 1-7.
- [31] McMichael, Anthony J., John W. Powles, Colin D. Butler, Ricardo Uauy. Food, Livestock Production, Energy, Climate Change, and Health. Lancet, 2007, 370 (9594): 1253-63. https://doi.org/10.1016/S0140-6736(07)61256-2
- [32] Martin, C., D. P. Morgavi, M. Doreau. Methane Mitigation in Ruminants: From Microbe to the Farm Scale. Animal, 2010, 4(3): 351-65. https://doi.org/10.1017/ S1751731 109990620
- [33] Meale, S. J., T. A. McAllister, K. A. Beauchemin, O. M. Harstad, A. V. Chaves. Strategies to Reduce Greenhouse Gases from Ruminant Livestock. Acta Agriculturae Scandinavica A: Animal Sciences, 2012, 62 (4): 199-211. https:// doi.org/ 10.1080/ 09064702
- [34] VAN SOEST, P.J.. Nutritional ecology of the ruminant. 2.ed. New York: Cornell University Press, 1994: 476.
- [35] Beauchemin KA, Kreuzer M, O'Mara F, McAllister TA. Nutritional management for enteric methane abatement: a review. Australian Journal of Experimental Agriculture, 2008, 48: 21-27.
- [36] ICF International. Greenhouse gas mitigation options and costs for agricultural land and animal production within the United States, 2013. http://www. usda.gov/oce/ climate change/mitigation technologies/GHG Mitigation Options.pdf
- [37] Itabashi H.. Reducing ruminal methane production by chemical and biological manipulation. In Greenhouse Gases and Animal Agriculture, 2002: 139-144 [J Takahasi and BA Young, editors]. Amsterdam: Elsevier Science.
- [38] Wood JM, Kennedy FS, Wolfe RS. The reaction of multi-halogenated hydrocarbons with free and bound reduced vitamin B12. Biochemistry, 1968, 7: 1707-1713.
- [39] Tomkins N., Hunter R.. Methane reduction in beef cattle using a novel antimethanogen. Anim Prod Aust, 2004, 25: 329.
- [40] Goel, Gunjan, Harinder P S Makkar, Klaus Becker. Inhibition of Methanogens by Bromochloromethane: Effects on Microbial Communities and Rumen Fermentation Using Batch and Continuous Fermentations, 2009.

https://doi.org/10.1017/S0007114508076198

[41] Lascano, Carlos E., Edgar Cárdenas. Alternatives for Mitigation of Methane Emission in Livestock Systems. Brazilian Journal of Animal Science, 2010, 39 (SUPPL. 1): 175-82. (in Portuguese) https://doi.org/10.1590/S1516-35982010001300020

- [42] Kumar, Anil, Puniya Guru, Angad Dev.. Controlling Methane Emissions from Ruminants Employing Bacteriocin Controlling Methane Emissions from Ruminants, no. December, 2013: 140-53. https://doi.org/10.13140/RG.2.1.3520.2401
- [43] Klieve, A.V., Joblin, K.. Comparison in hydrogen utilisation of ruminal and marsupial reductive acetogens. In R. Kennedy, (eds) 5 Year Research Progress Report 2002 - 2007, The Pastoral Greenhouse Gas Research Consortium, Wellington, New Zealand,

2007: 34 - 35.

- [44] Powell, J. M., M. J. Aguerre, M. A. Wattiaux. Dietary crude protein and tannin impact dairy manure chemistry and ammonia emissions from incubated soils. J. Environ. Qual. 2011a, 40: 1767-1774.
- [45] Hatch, D., H. Trindade, L. Cardenas, J. Carneiro, J. Hawkins, D. Scholefield, D. Chadwick. Laboratory Study of the Effects of Two Nitrification Inhibitors on Greenhouse Gas Emissions from a Slurry-Treated Arable Soil: Impact of Diurnal Temperature Cycle. Biology and Fertility of Soils, 2005, 41(4): 225-32. https://doi.org/10.1007/s00374-005-0836-9

# **Author Guidelines**

This document provides some guidelines to authors for submission in order to work towards a seamless submission process. While complete adherence to the following guidelines is not enforced, authors should note that following through with the guidelines will be helpful in expediting the copyediting and proofreading processes, and allow for improved readability during the review process.

#### I. Format

- Program: Microsoft Word (preferred)
- Font: Times New Roman
- Size: 12
- Style: Normal
- Paragraph: Justified
- Required Documents

#### **II** . Cover Letter

All articles should include a cover letter as a separate document.

The cover letter should include:

• Names and affiliation of author(s)

The corresponding author should be identified.

Eg. Department, University, Province/City/State, Postal Code, Country

• A brief description of the novelty and importance of the findings detailed in the paper

#### Declaration

v Conflict of Interest

Examples of conflicts of interest include (but are not limited to):

- Research grants
- Honoria
- Employment or consultation
- Project sponsors
- Author's position on advisory boards or board of directors/management relationships
- Multiple affiliation
- Other financial relationships/support
- Informed Consent

This section confirms that written consent was obtained from all participants prior to the study.

• Ethical Approval

Eg. The paper received the ethical approval of XXX Ethics Committee.

- Trial Registration
- Eg. Name of Trial Registry: Trial Registration Number

#### • Contributorship

The role(s) that each author undertook should be reflected in this section. This section affirms that each credited author has had a significant contribution to the article.

1. Main Manuscript

2. Reference List

3. Supplementary Data/Information

Supplementary figures, small tables, text etc.

As supplementary data/information is not copyedited/proofread, kindly ensure that the section is free from errors, and is presented clearly.

#### **Ⅲ**. Abstract

A general introduction to the research topic of the paper should be provided, along with a brief summary of its main results and implications. Kindly ensure the abstract is self-contained and remains readable to a wider audience. The abstract should also be kept to a maximum of 200 words.

Authors should also include 5-8 keywords after the abstract, separated by a semi-colon, avoiding the words already used in the title of the article.

Abstract and keywords should be reflected as font size 14.

#### **W.** Title

The title should not exceed 50 words. Authors are encouraged to keep their titles succinct and relevant.

Titles should be reflected as font size 26, and in bold type.

#### **IV. Section Headings**

Section headings, sub-headings, and sub-subheadings should be differentiated by font size.

Section Headings: Font size 22, bold type Sub-Headings: Font size 16, bold type Sub-Subheadings: Font size 14, bold type Main Manuscript Outline

#### V. Introduction

The introduction should highlight the significance of the research conducted, in particular, in relation to current state of research in the field. A clear research objective should be conveyed within a single sentence.

#### **VI.** Methodology/Methods

In this section, the methods used to obtain the results in the paper should be clearly elucidated. This allows readers to be able to replicate the study in the future. Authors should ensure that any references made to other research or experiments should be clearly cited.

#### **W**. Results

In this section, the results of experiments conducted should be detailed. The results should not be discussed at length in

this section. Alternatively, Results and Discussion can also be combined to a single section.

#### **W**. Discussion

In this section, the results of the experiments conducted can be discussed in detail. Authors should discuss the direct and indirect implications of their findings, and also discuss if the results obtain reflect the current state of research in the field. Applications for the research should be discussed in this section. Suggestions for future research can also be discussed in this section.

#### IX. Conclusion

This section offers closure for the paper. An effective conclusion will need to sum up the principal findings of the papers, and its implications for further research.

#### X. References

References should be included as a separate page from the main manuscript. For parts of the manuscript that have referenced a particular source, a superscript (ie. [x]) should be included next to the referenced text.

[x] refers to the allocated number of the source under the Reference List (eg. [1], [2], [3])

In the References section, the corresponding source should be referenced as:

[x] Author(s). Article Title [Publication Type]. Journal Name, Vol. No., Issue No.: Page numbers. (DOI number)

#### XI. Glossary of Publication Type

J = Journal/Magazine

- M = Monograph/Book
- C = (Article) Collection
- D = Dissertation/Thesis
- P = Patent
- S = Standards
- N = Newspapers
- R = Reports

Kindly note that the order of appearance of the referenced source should follow its order of appearance in the main manuscript.

Graphs, Figures, Tables, and Equations

Graphs, figures and tables should be labelled closely below it and aligned to the center. Each data presentation type should be labelled as Graph, Figure, or Table, and its sequence should be in running order, separate from each other. Equations should be aligned to the left, and numbered with in running order with its number in parenthesis (aligned right).

#### XII. Others

Conflicts of interest, acknowledgements, and publication ethics should also be declared in the final version of the manuscript. Instructions have been provided as its counterpart under Cover Letter.

#### About the Publisher

Bilingual Publishing Co. (BPC) is an international publisher of online, open access and scholarly peer-reviewed journals covering a wide range of academic disciplines including science, technology, medicine, engineering,education and social science. Reflecting the latest research from a broad sweep of subjects, our content is accessible worldwide – both in print and online.

BPC aims to provide an analytics as well as platform for information exchange and discussion that help organizations and professionals in advancing society for the betterment of mankind. BPC hopes to be indexed by well-known databases in order to expand its reach to the science community, and eventually grow to be a reputable publisher recognized by scholars and researchers around the world.

BPC adopts the Open Journal Systems, see on http://ojs.bilpublishing.com



Google Scholar

# **Database Inclusion**



Creative Commons



Crossref



MyScienceWork



Tel:+65 65881289 E-mail:contact@bilpublishing.com Website:www.bilpublishing.com